

The use of genomic and gene expression large-scale data for the analyses of sexual evolution

Maria D. Vibranovski



Instituto de
biociências

UNIVERSIDADE DE SÃO PAULO
INSTITUTO DE BIOCIÊNCIAS
Departamento de Genética e Biologia Evolutiva



15 de abril de 2015 - UFRJ

Acknowledgments



Manyuan Long
Department of Ecology and Evolution
The University of Chicago



Timothy L. Karr
Drosophila Genetic Resource Center
Kyoto Institute of Technology



Hedibert F. Lopes
INSPER – São Paulo

Outline

1. Background in Sexual Evolution

2. Biological Problem

3. Large-scale Data

4. Statistical Approaches

5. Ongoing Biological Problems

6. Perspectives

1. Background in Sexual Evolution

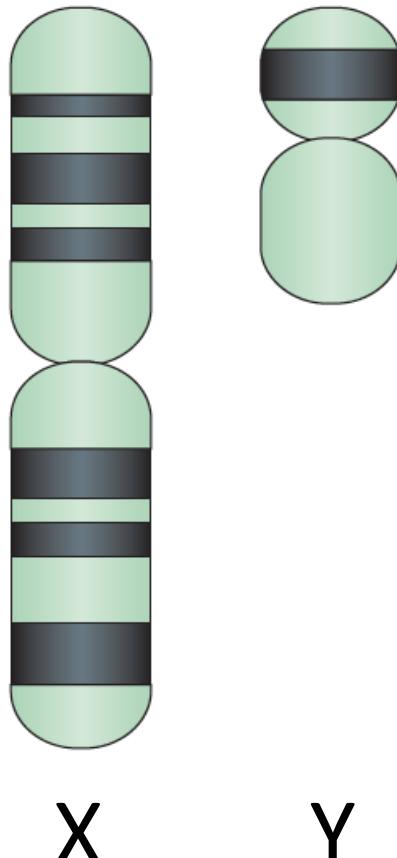
Why are there phenotypic differences between sexes?



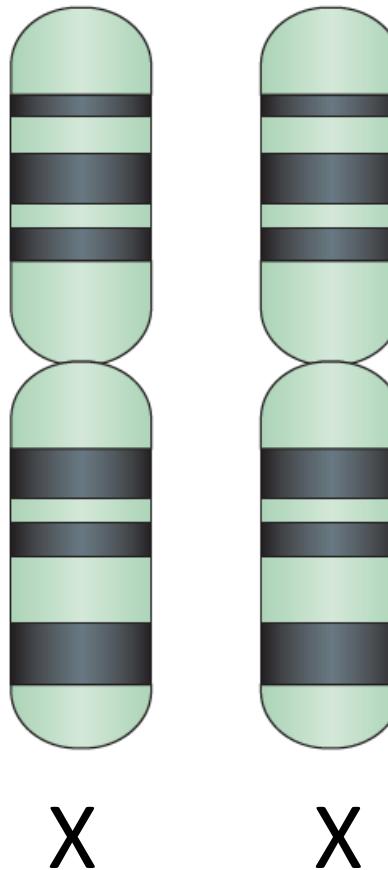
1. Background in Sexual Evolution

Sex Chromosomes

Male



Female



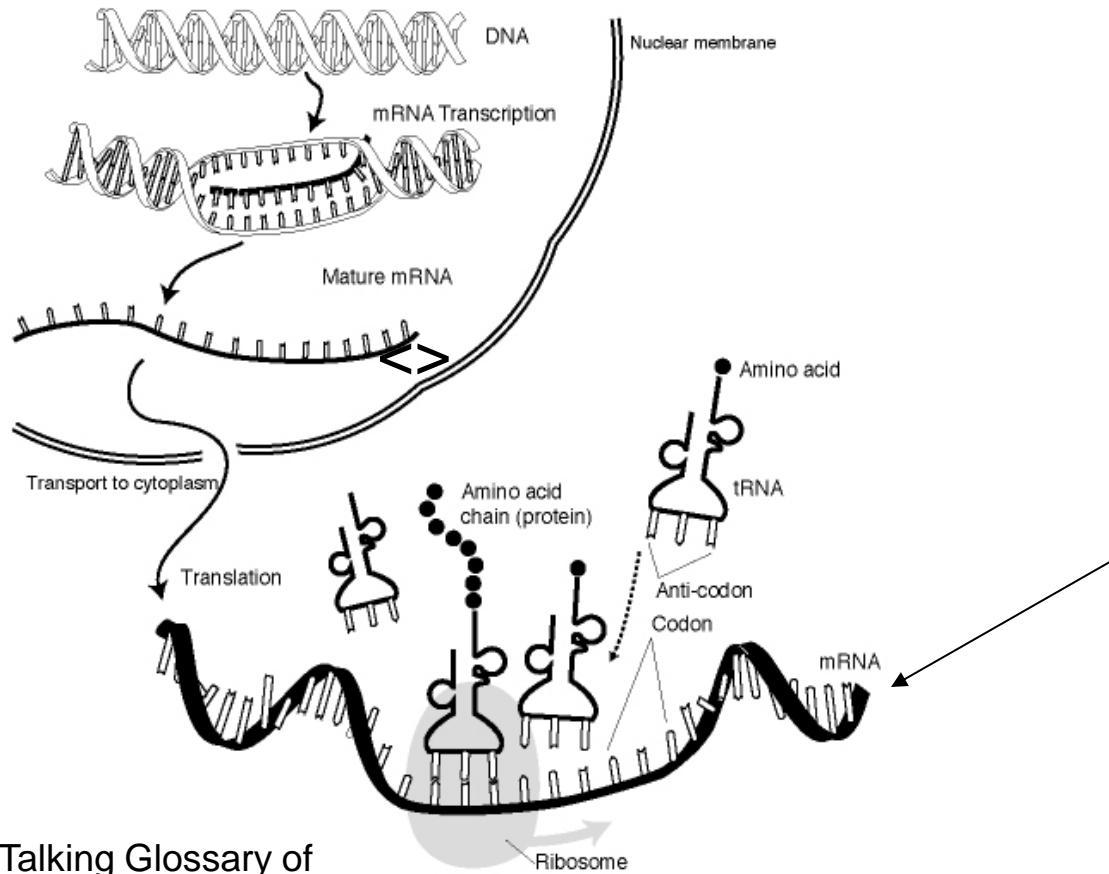
X

Y

X

X

1. Background in Sexual Evolution



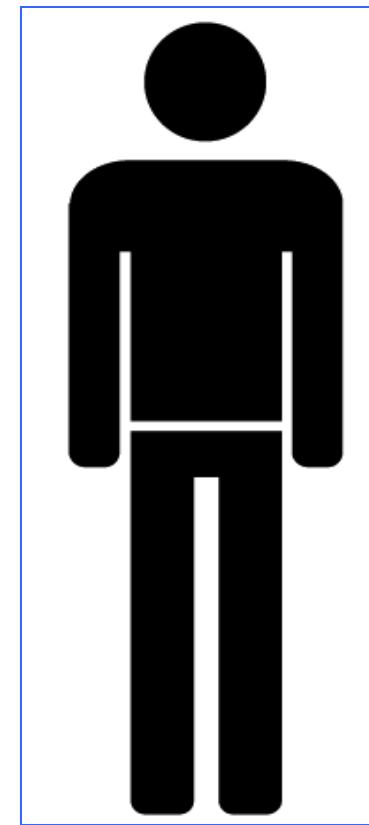
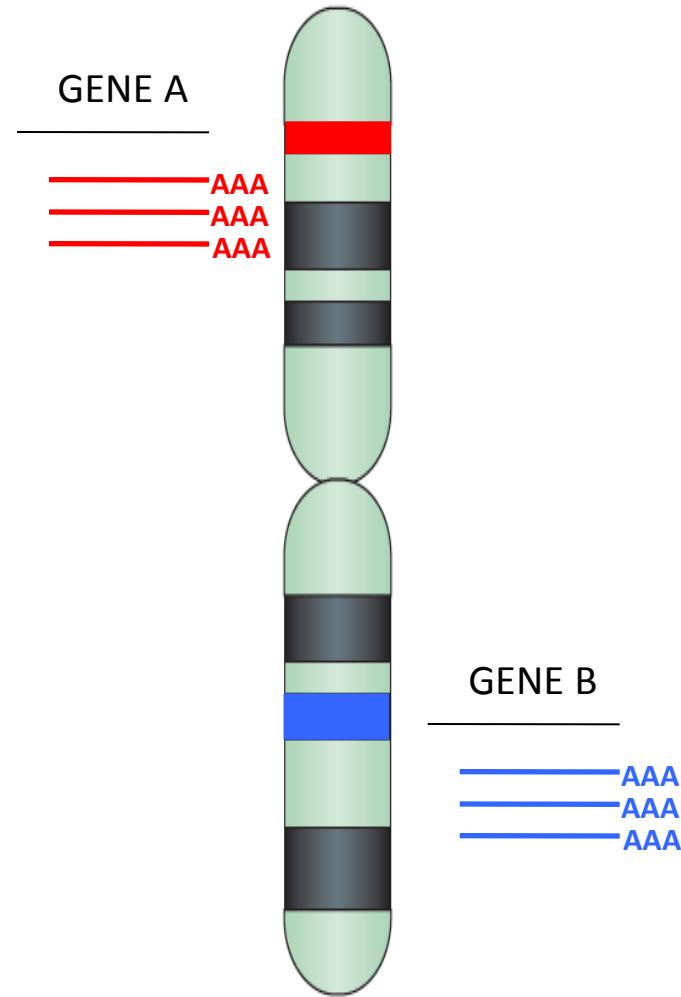
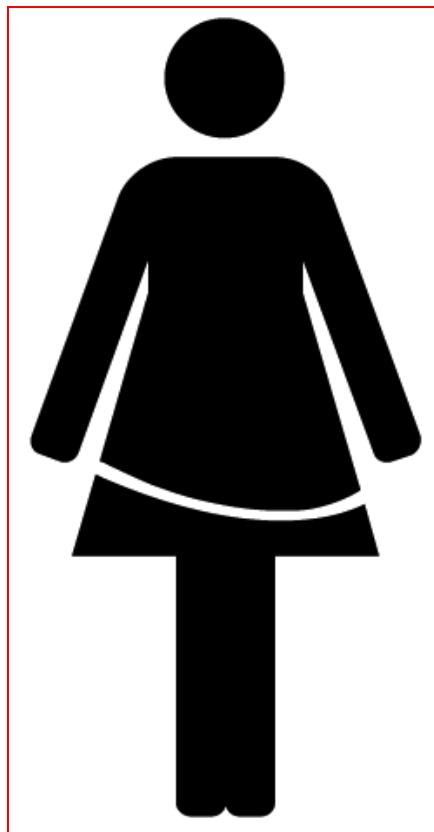
From Talking Glossary of Genetics

Measure the quantity of messenger RNA (mRNA) in the cell

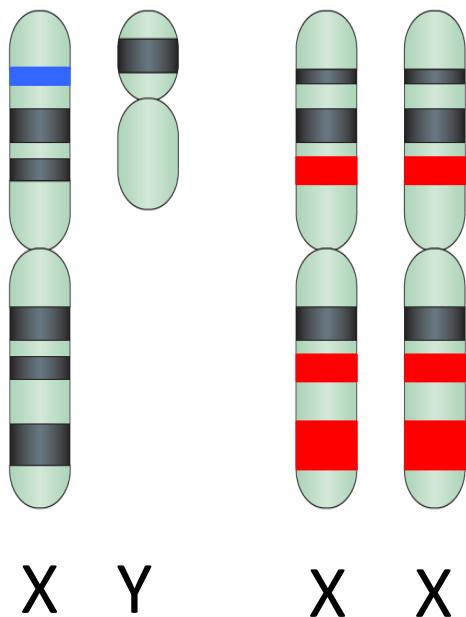
Gene activity = gene expression = number of mRNA molecules

1. Background in Sexual Evolution

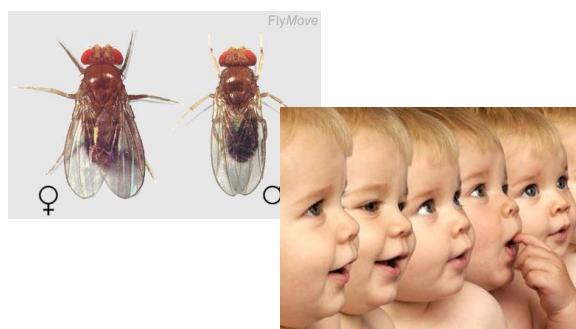
Sex-biased Gene Expression



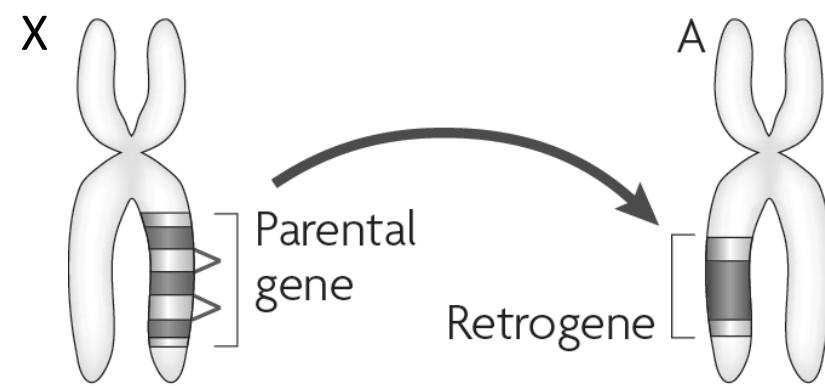
2. Biological Problem



What molecular mechanisms and genetics processes are involved?



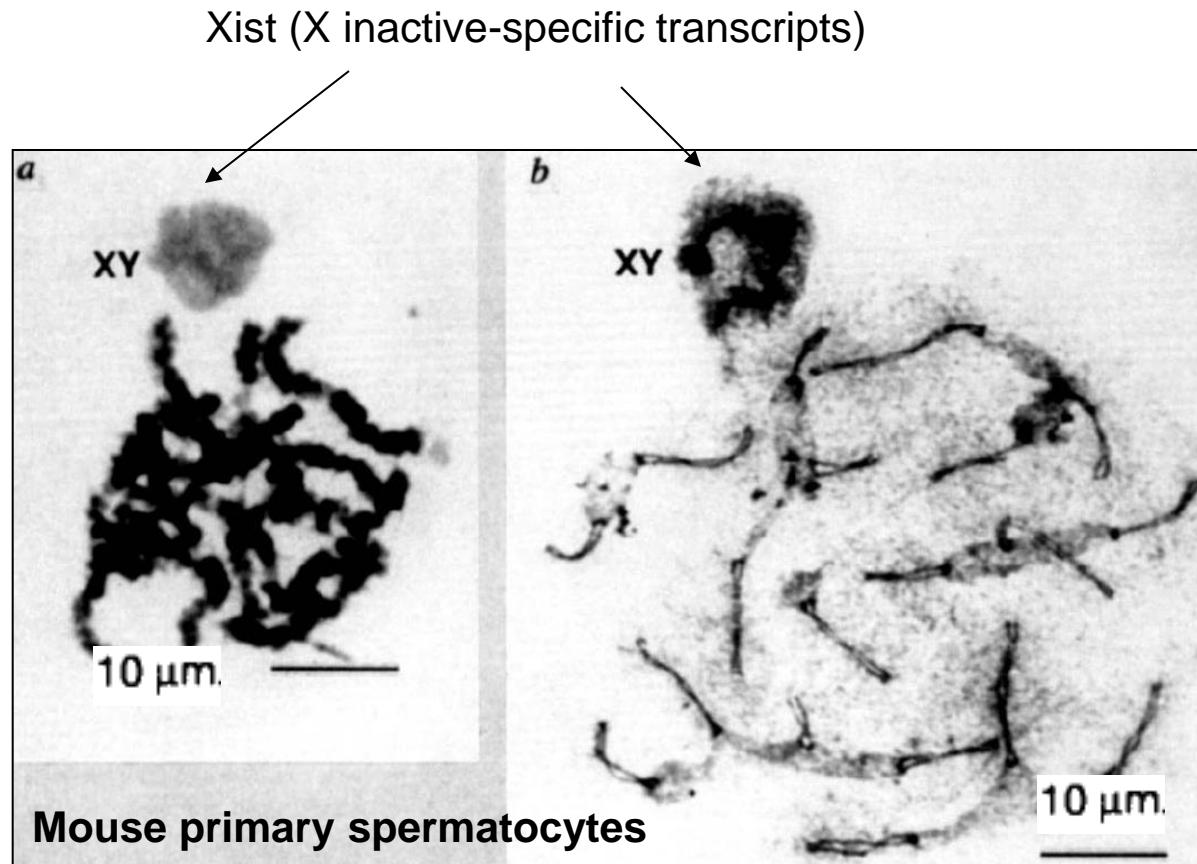
XY systems



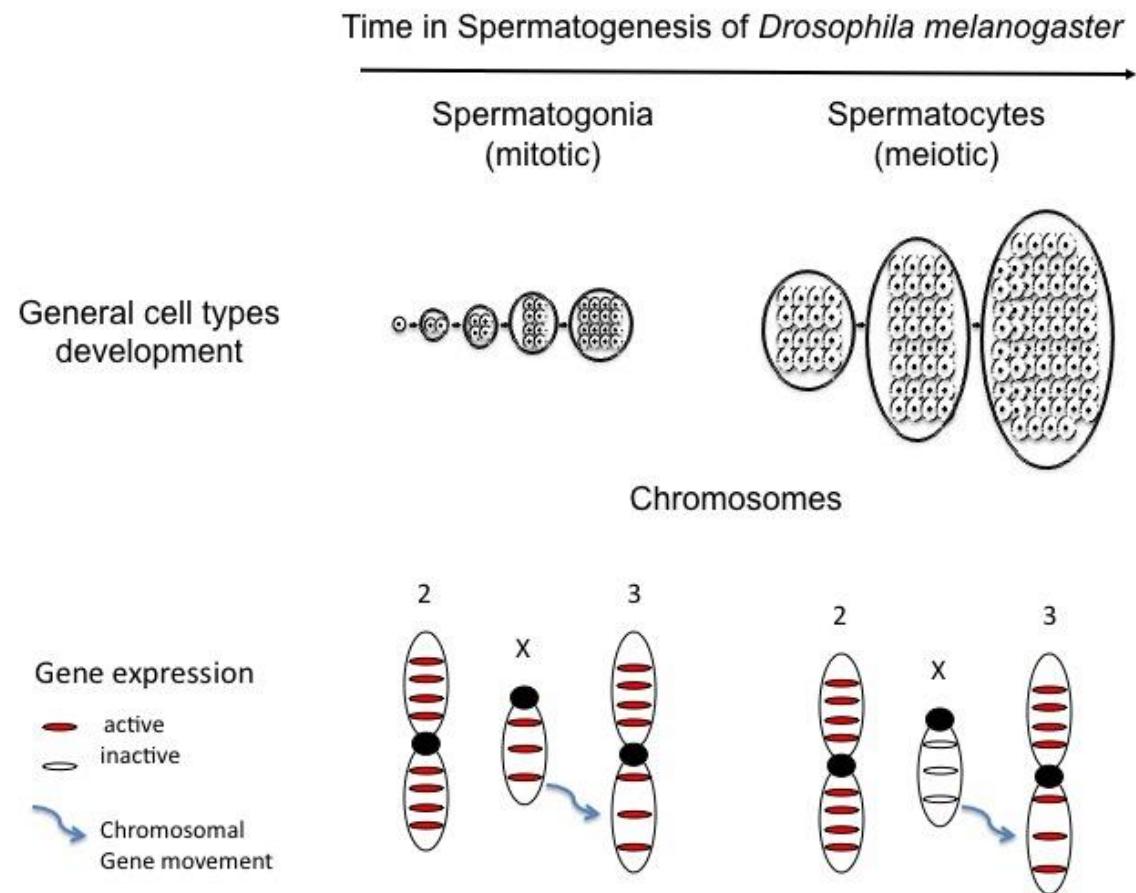
Testis bias

2. Biological Problem

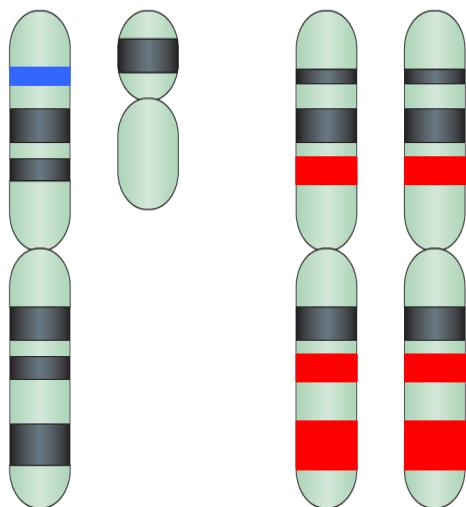
Meiotic Sex Chromosome Inactivation (MSCI)



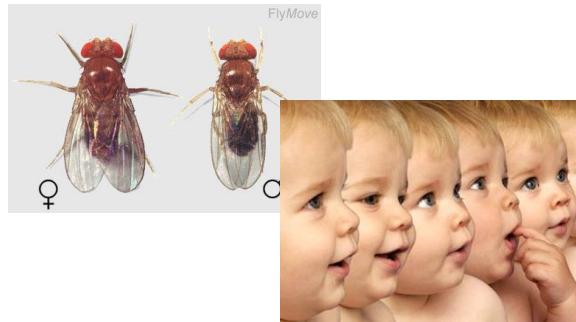
2. Biological Problem



2. Biological Problem

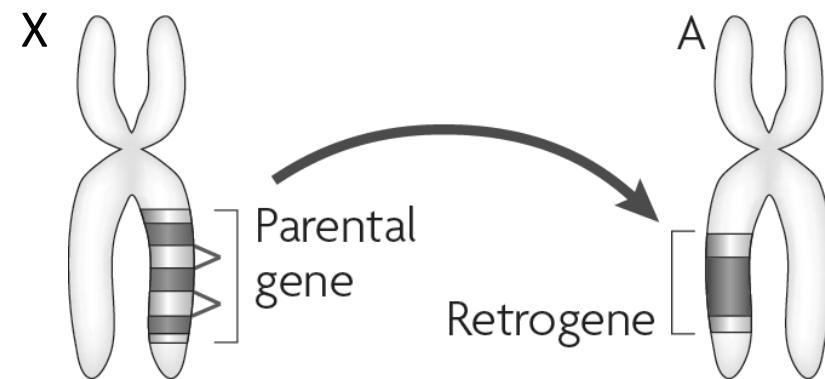


X Y X X



XY systems

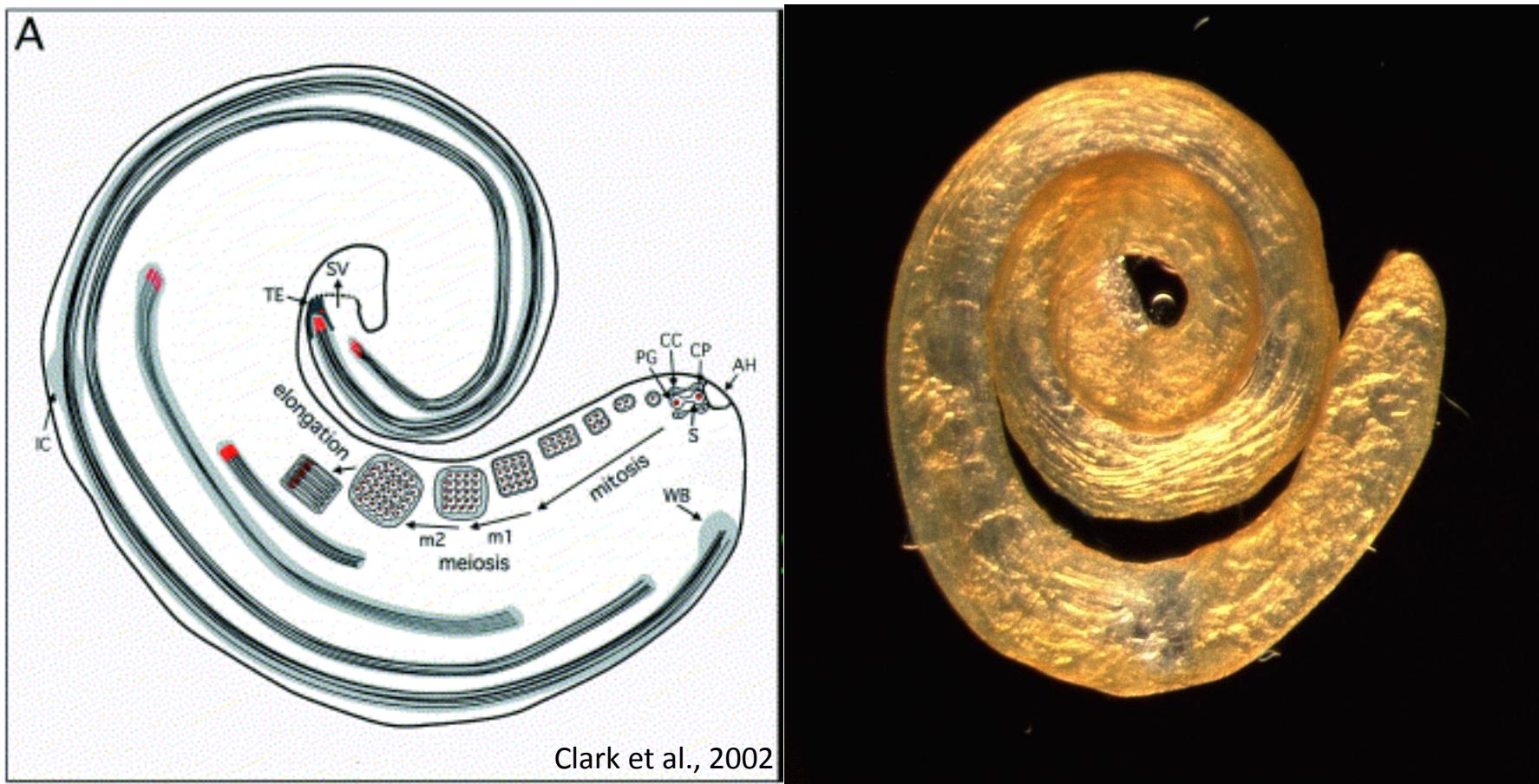
- Does MSCI exist in *Drosophila*?
- Is MSCI involved in the distribution of sex-biased genes in the genome?



Testis bias

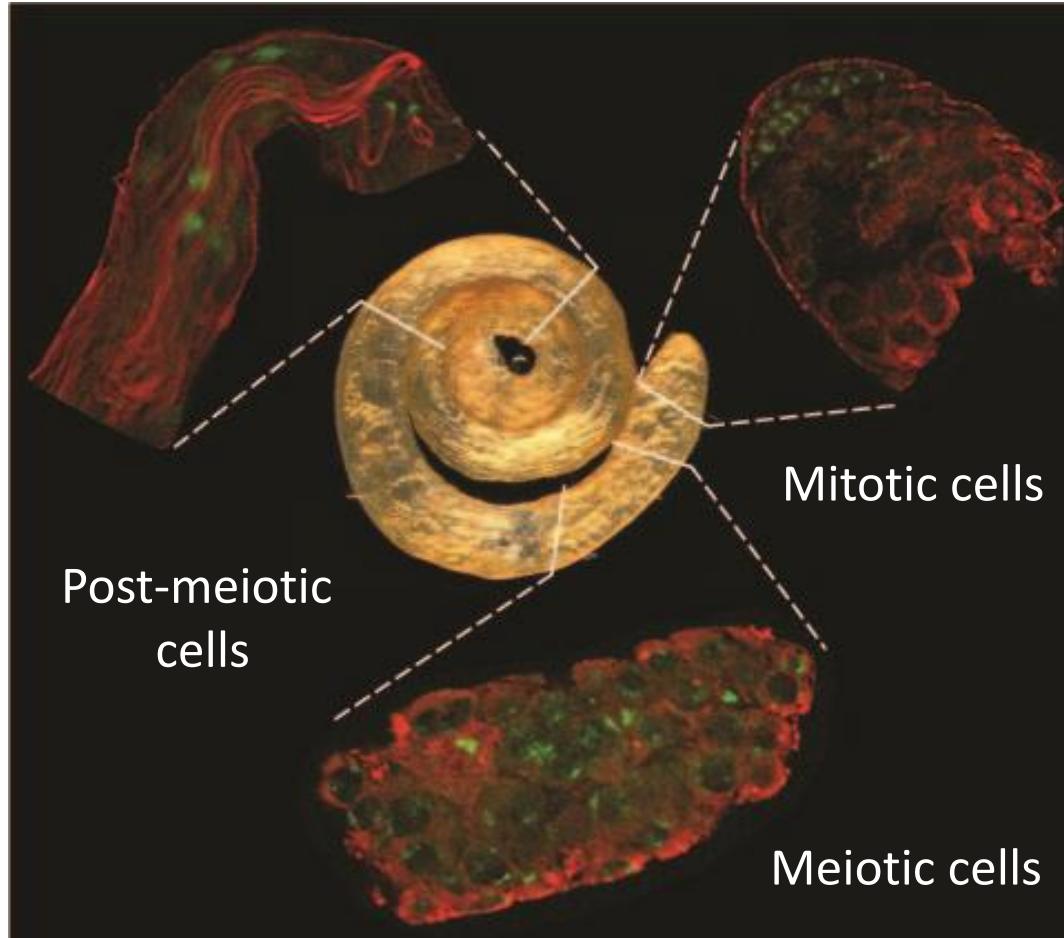
3. Large-scale Data

Drosophila melanogaster Spermatogenesis



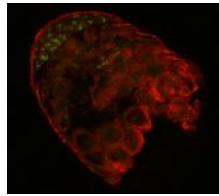
3. Large-scale Data

Isolation of Spermatogenic Cells



3. Large-scale Data

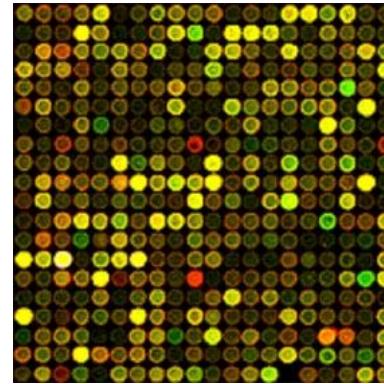
Tissue Isolation (n=3)



RNA
extraction



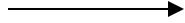
Microarray
Hybridization



3. Large-scale Data

How to count the number of mRNA molecules for each gene in a cell?

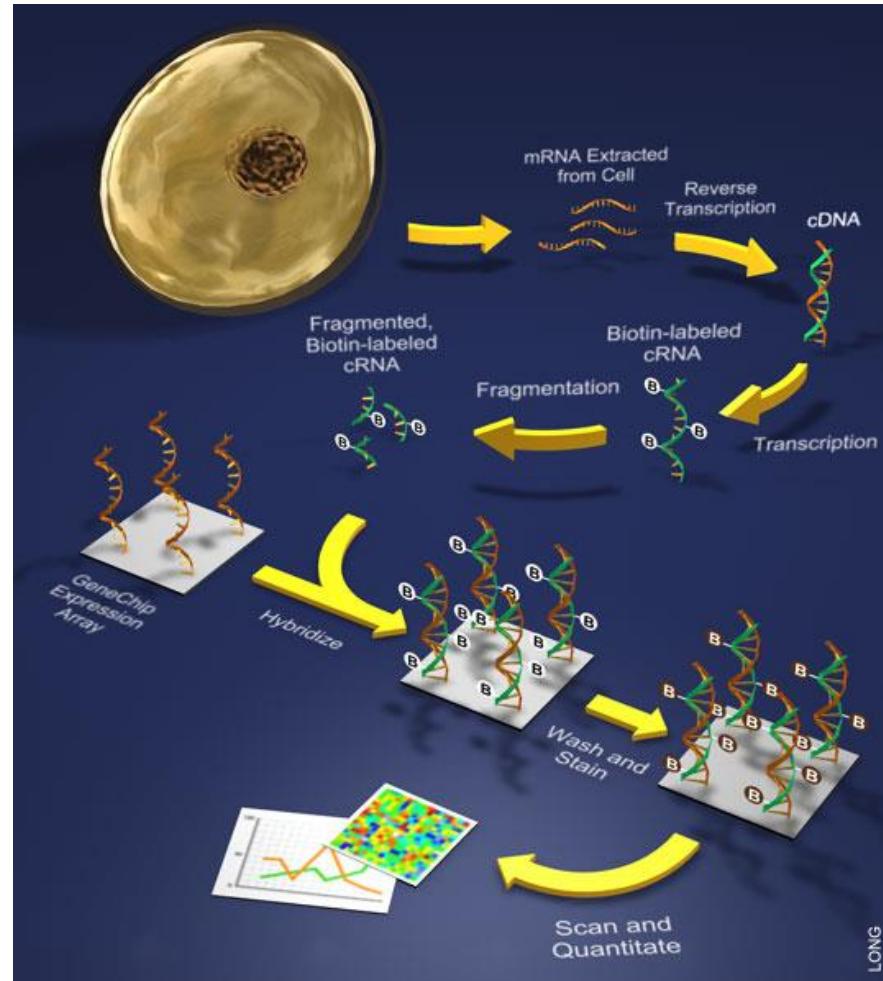
Microarray: A chip containing small pieces of DNA corresponding to all *Drosophila* genes



3. Large-scale Data

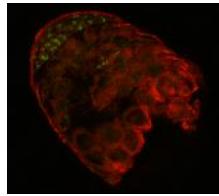
Microarray

Specific case for gene expression measure: the GeneChip array



3. Large-scale Data

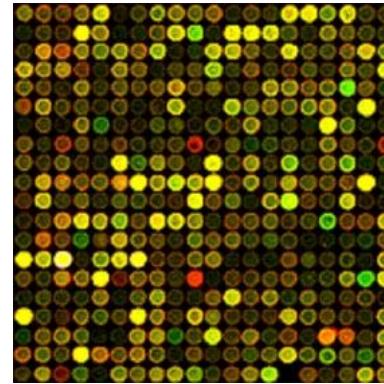
Tissue Isolation (n=3)



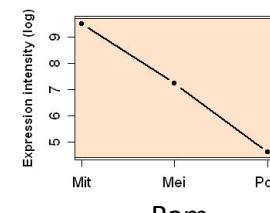
RNA
extraction



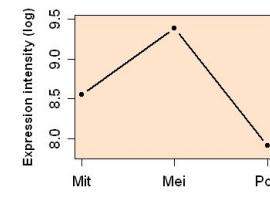
Microarray
Hybridization



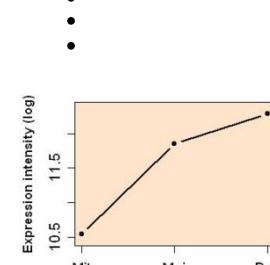
Gene profile



Bam



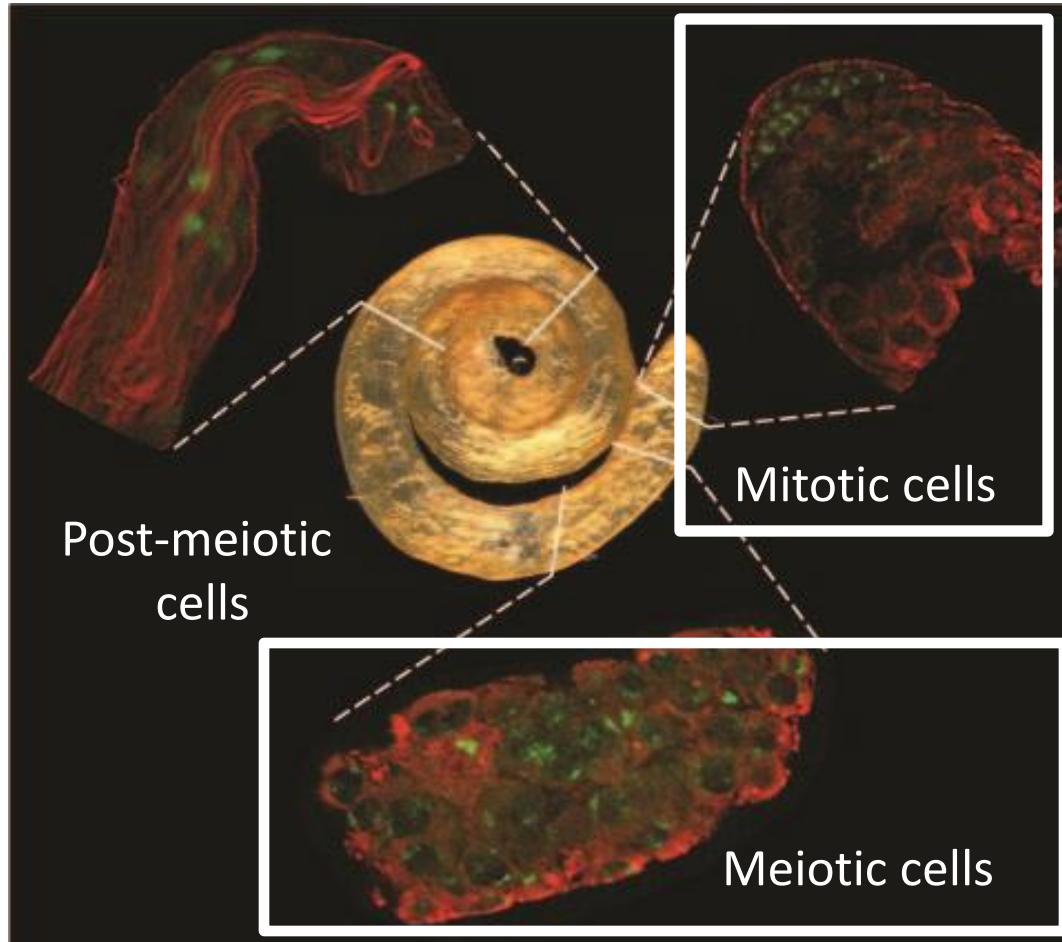
ms(3)K81



Mst35Bb

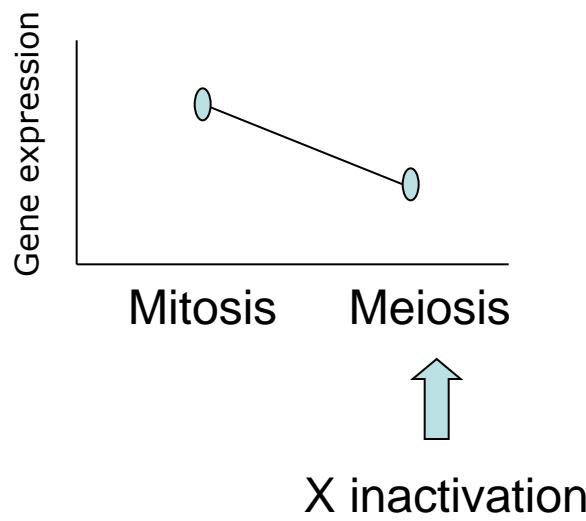
4. Statistical Approaches

Meiotic Sex Chromosome Inactivation (MSCI)



4. Statistical Approaches

X inactivation occurs when the difference in activity (meiosis-mitosis) in X is lower than the difference in activity in autosome.



$$\theta = \text{gene expression /activity}$$

$$\theta_{\text{meiosis}}^X - \theta_{\text{mitosis}}^X < \theta_{\text{meiosis}}^A - \theta_{\text{mitosis}}^A$$

4. Statistical Approaches

Gene intensity

2982 genes (X chromosomes)

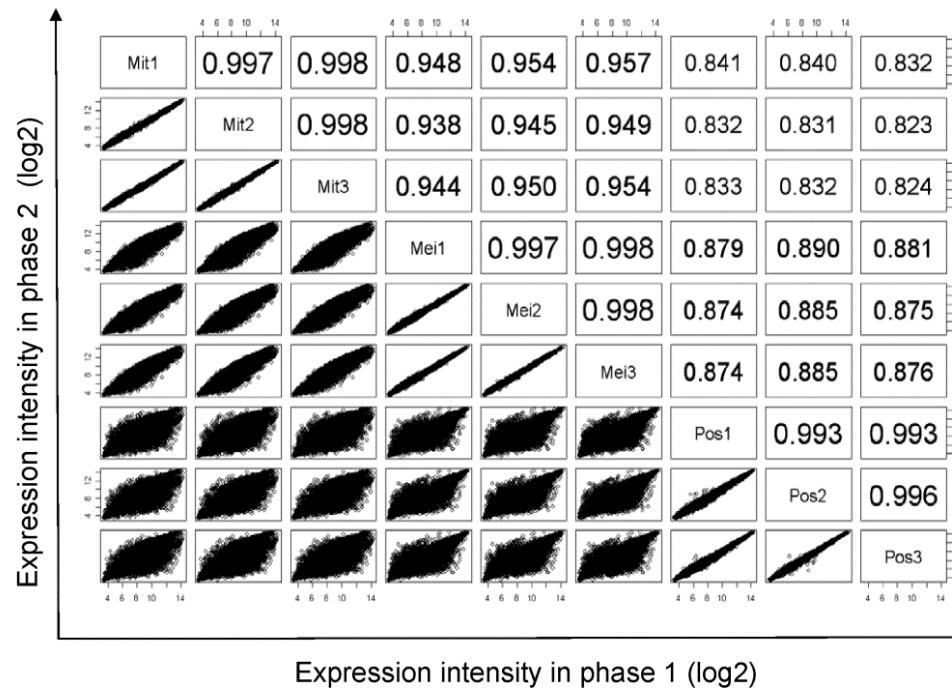
15099 genes (autosomes)

Spermatogenic replicates

3 mitotic

3 meiotic

3 post-meiotic



Gene	Mit1	Mit2	Mit3	Mei1	Mei2	Mei3
292	10.182	10.199	10.395	9.798	9.880	9.862
792	5.273	5.357	5.509	5.548	5.577	5.587
1966	5.924	5.914	5.945	6.297	6.779	6.643
2811	5.177	5.197	5.168	5.092	5.166	5.169

Gene	Mit1	Mit2	Mit3	Mei1	Mei2	Mei3
2869	9.884	9.872	9.758	11.468	11.174	11.213
3939	4.833	4.755	4.775	4.827	4.839	4.804
10541	4.932	4.932	4.969	5.073	4.876	4.986
12928	10.331	10.485	10.524	11.480	11.628	11.514

4. Statistical Approaches

For chromosome i (X or A) and gene l , 3 replicates are measured

Mitosis: $mit_{i1l}, mit_{i2l}, mit_{i3l}$ $E(mit_{ikl}) = \theta_{il}^{mit}$

Meiosis: $mei_{i1l}, mei_{i2l}, mei_{i3l}$ $E(mei_{ikl}) = \theta_{il}^{mei}$

The objective is to learn whether genes are

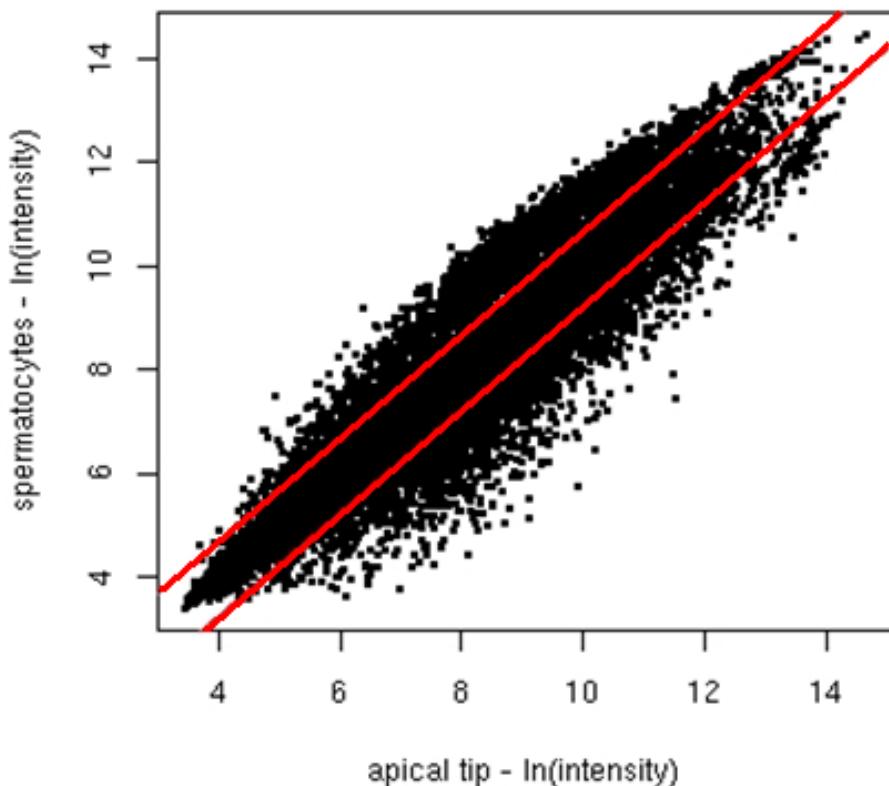
Differently expressed: $\begin{cases} H_{1,il} : \theta_{il}^{mei} > \theta_{il}^{mit} & (\text{OVER}) \\ H_{2,il} : \theta_{il}^{mei} < \theta_{il}^{mit} & (\text{UNDER}) \end{cases}$

or

Equally expressed: $H_{3,il} : \theta_{il}^{mei} = \theta_{il}^{mit}$ (EQUAL).

4. Statistical Approaches

How do biologists usually approach this problem?



APPROACH 1:

Identify differently expressed genes based on 2 fold intensity differences (based on homotypic experiments);

APPROACH 2:

Identify differently expressed genes by controlling type I error;

APPROACH 3:

Identify differently expressed genes by controlling false discovery rates;

APPROACH 4:

Hierarchical Bayes.

4. Statistical Approaches

How do biologists usually approach this problem?

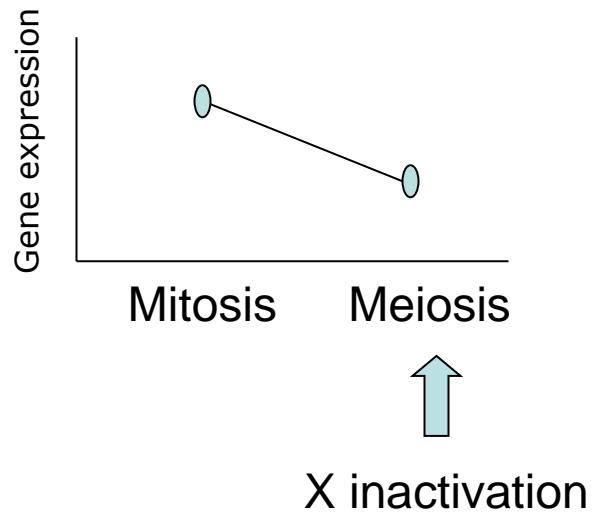
APPROACH 1, 2 or 3
Identify differently expressed genes



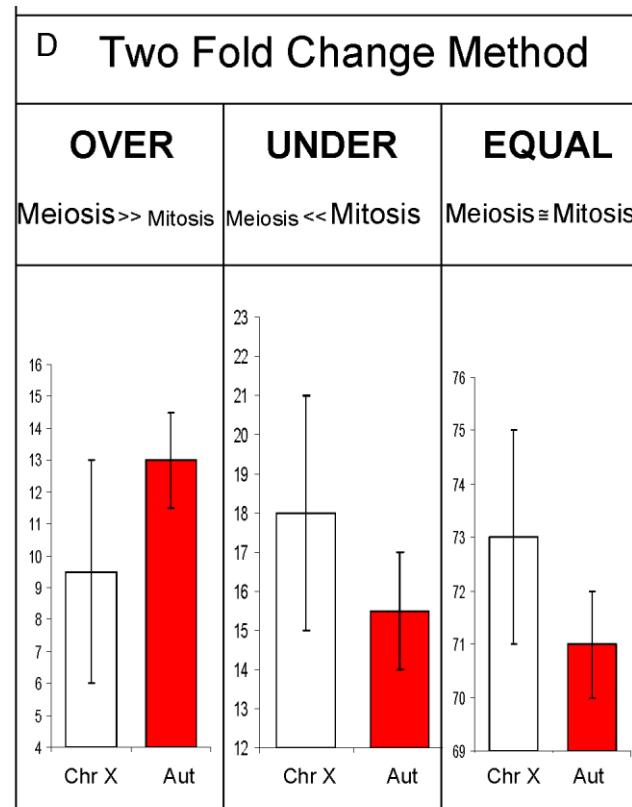
Counts

Expression	ChrX	ChrA
Under expressed	537(18%)	2416(16%)
Equally expressed	2147(72%)	10720(71%)
Over expressed	298(10%)	1963(13%)
Total	2982	15099

4. Statistical Approaches



Proportion of genes (%)



4. Statistical Approaches

Approach 2: *False positive rate (FPR)*

	(E) Evidence against H	(NE) No evidence against H	Total
(A) Over/under expressed	τ	$\eta_1 - \tau$	η_1
(H) Equally expressed	η	$\eta_0 - \eta$	η_0
Total	x	n-x	n

Measures error rate (evidence against H) when H is true: $E(\eta/\eta_0)$

$$FPR=0.05 \Rightarrow E(\eta) < 0.05E(\eta_0)$$

Bonferroni correction

When evidence is measured by $\{p\text{-value} < 0.05/n\}$, then

$$\Pr(\eta > 0) < 0.05$$

Controlling $E(\eta) < 0.05n$ is too liberal!

Controlling $\Pr(\eta > 0) < 0.05$ is too conservative!

4. Statistical Approaches

Approach 3: *False discovery rate (FDR)*

	(E) Evidence against H	(NE) No evidence against H	Total
(A) Over/under expressed	τ	$\eta_1 - \tau$	η_1
(H) Equally expressed	η	$\eta_0 - \eta$	η_0
Total	x	$n-x$	n

$$FDR(\alpha) = E\left(\frac{\eta(\alpha)}{x(\alpha)}\right) \approx \frac{E(\eta(\alpha))}{E(x(\alpha))} = \frac{\eta_0 \alpha}{E(x(\alpha))} \approx \frac{n \hat{\pi}_0 \alpha}{\sum_{i=1}^n 1(p_i < \alpha)}$$

$$\hat{\pi}_0 = \lim_{\lambda \rightarrow 1} \frac{\sum_{i=1}^n 1(p_i > \lambda)}{n(1-\lambda)} \quad (\text{estimate of true nulls})$$

$$FDR(\alpha) \approx \frac{\pi_0 \Pr(E | H)}{\Pr(E)} = \Pr(H | E) \quad (\text{Looks Bayesian!!})$$

4. Statistical Approaches

Approach 4: mixture model for differences

Because σ_x^2 and σ_A^2 (within gene variability) in the hierarchical model are negligible when compared τ^2 (between genes variability), the next model we implemented is a mixture of normals model that accounts for the excess of intensities around zero:

Model:

$$d_l = (\bar{x}_l^{mei} - \bar{x}_l^{mit}) \sim \pi N(\theta_1, \tau_1^2) + (1 - \pi) N(\theta_2, \tau_2^2) \quad l = 1, \dots, n$$

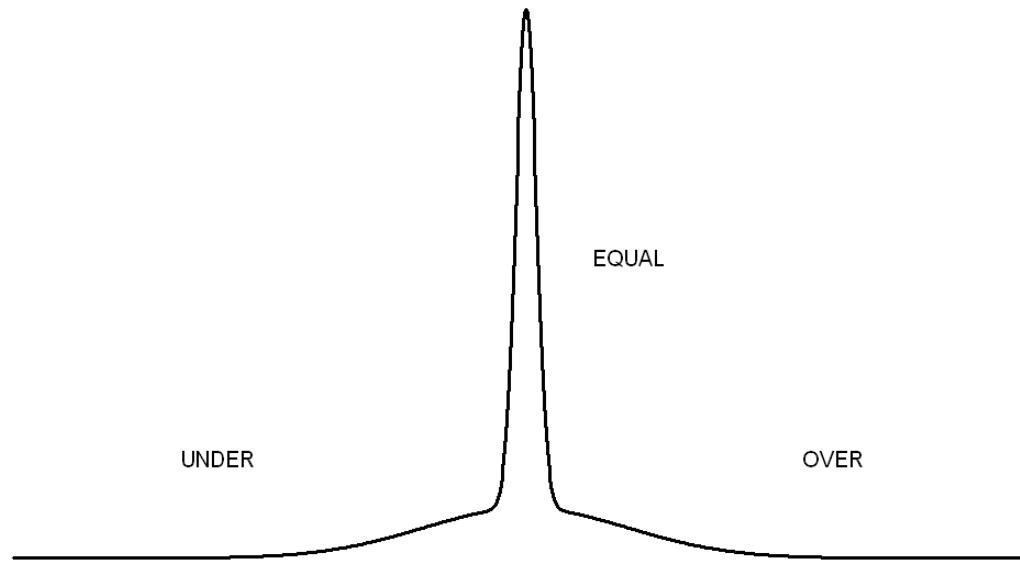
Prior:

$$p(\pi, \theta_1, \theta_2, \tau_1^2, \tau_2^2) \propto \tau_1^{-2} \tau_2^{-2}$$

4. Statistical Approaches

Approach 4: mixture model for differences

- Bayesian Mixture Model classifies genes as over or under expressed
 - Detect differential gene expression while comparing chromosomes distributions
 - Avoids the arbitrariness of folding
 - Avoids multiple testing distortions



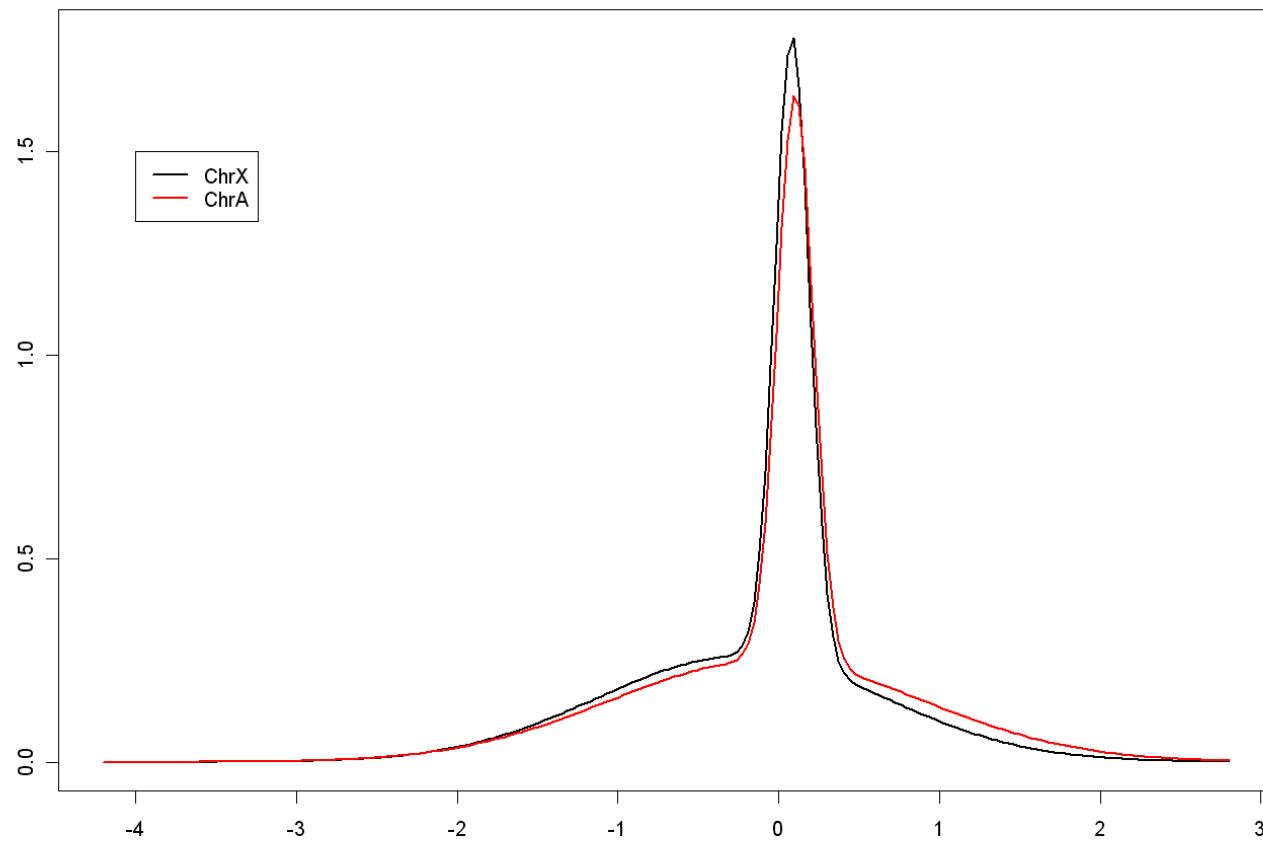
Lopes, Mueller and Rosner (2003)

4. Statistical Approaches

Approach 4: mixture model for differences

$$p(d | data) = \int (\pi f_N(d; \theta_1, \tau_1^2) + (1 - \pi) f_N(d; \theta_2, \tau_2^2)) p(\pi, \theta_1, \theta_2, \tau_1^2, \tau_2^2 | data) d\pi d\theta_1 d\theta_2 d\tau_1^2 d\tau_2^2$$

LOG(meiosis/Mitosis)



4. Statistical Approaches

Approach 4: mixture model for differences

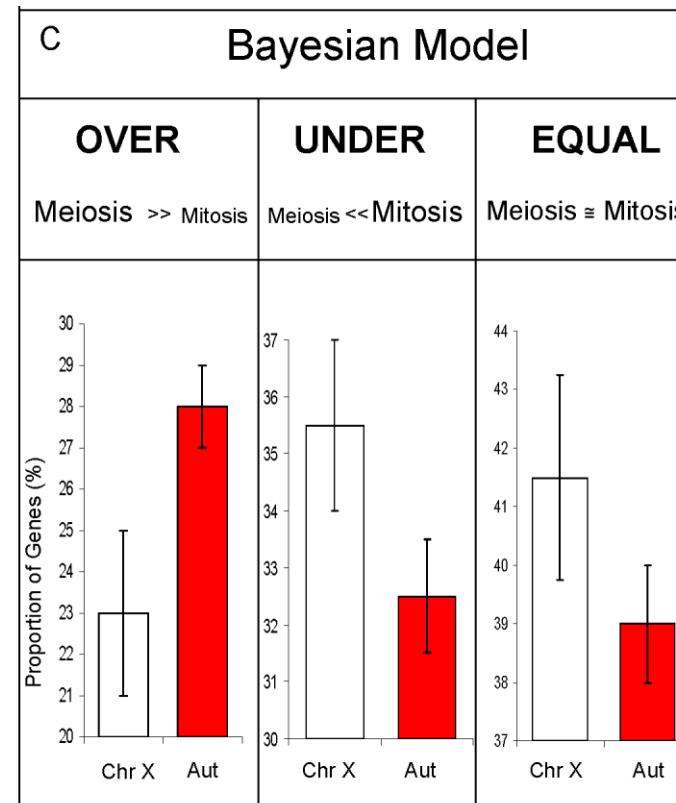
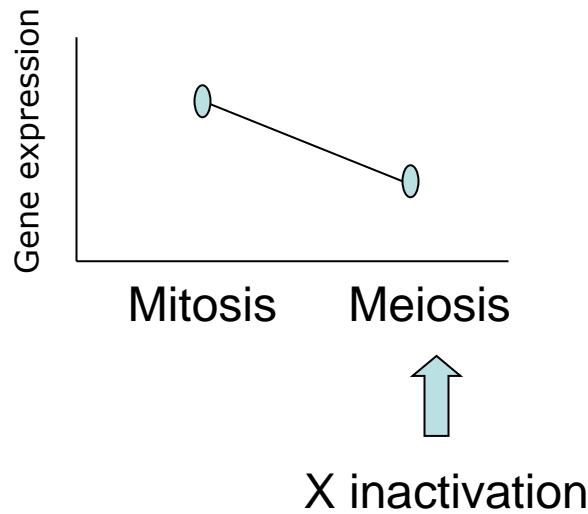
Classification (Prior): $Z_l \sim Ber(\pi)$

Classification (Posterior):

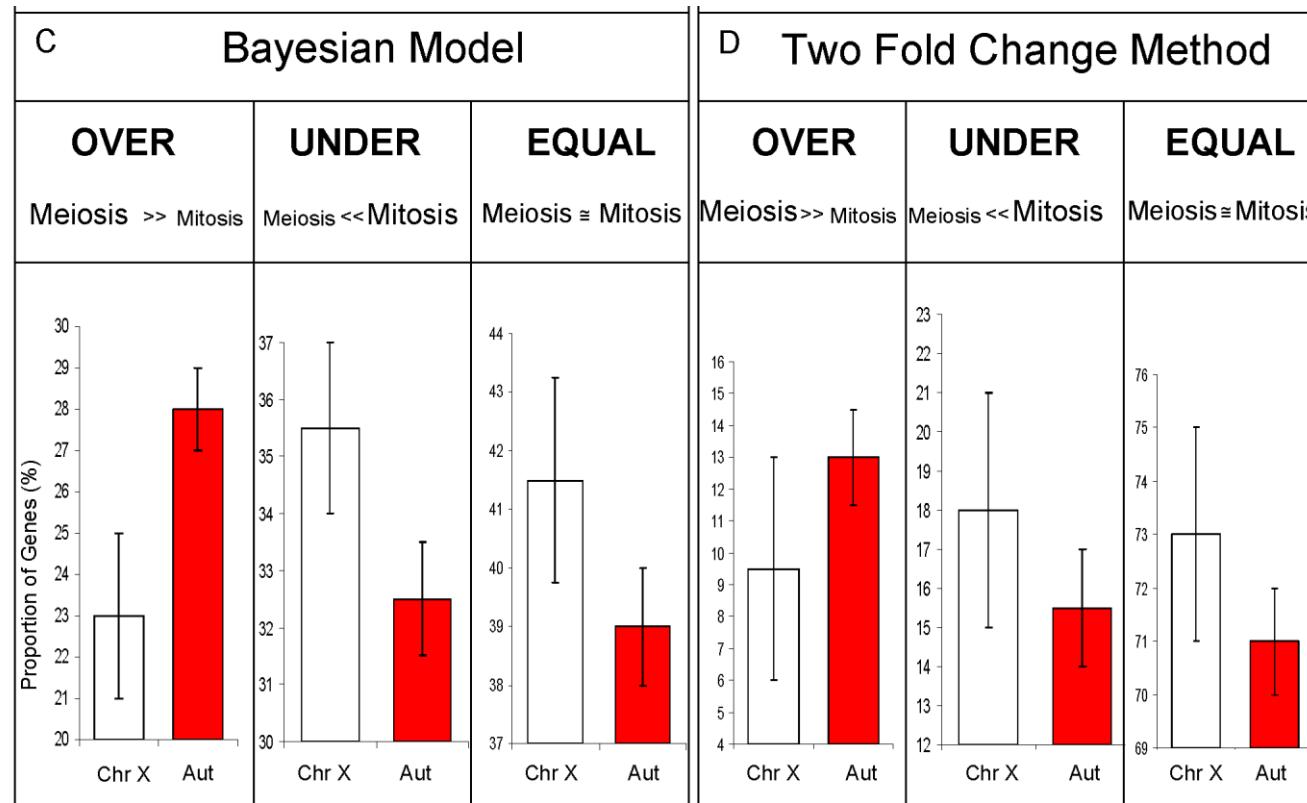
- $(Z_l = 1) \& (d_l > 0)$ – over expressed
- $(Z_l = 1) \& (d_l < 0)$ – under expressed
- $(Z_l = 0)$ – equally expressed

4. Statistical Approaches

Approach 4: Bayesian **mixture** model for differences

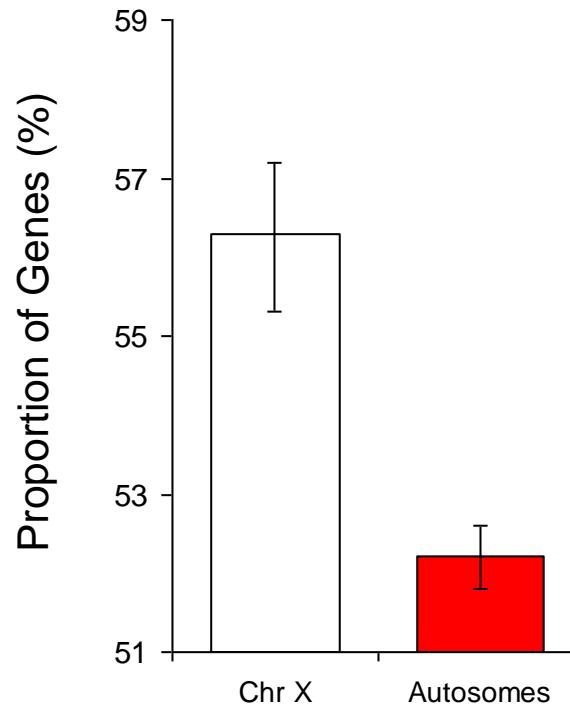
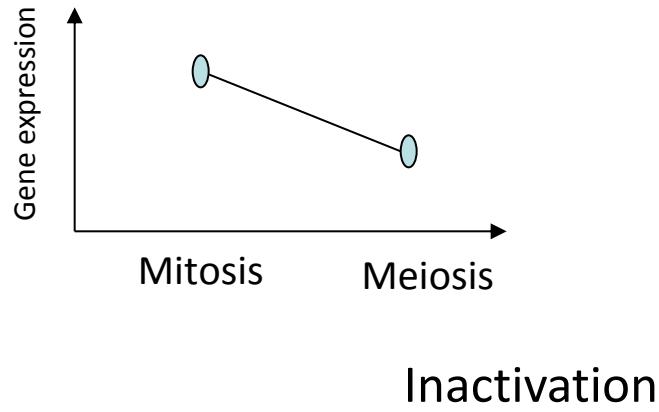


4. Statistical Approaches



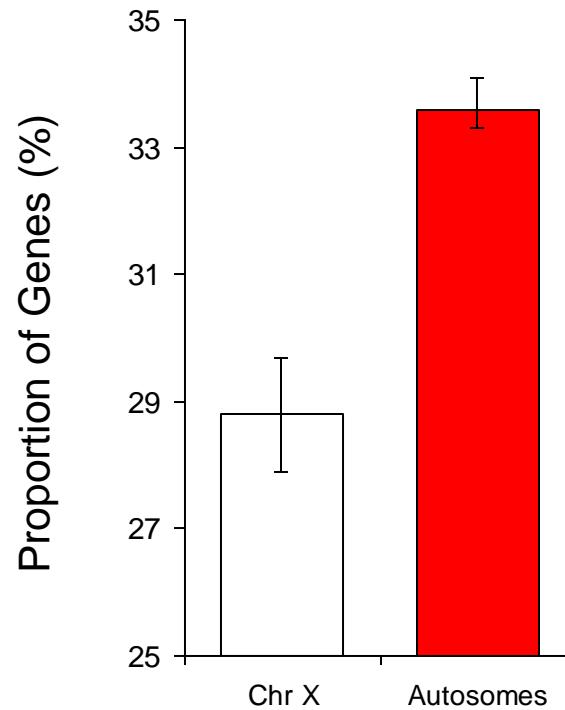
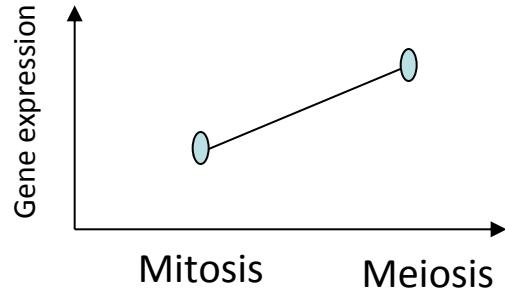
4. Statistical Approaches

Meiotic Sex Chromosome Inactivation (MSCI)

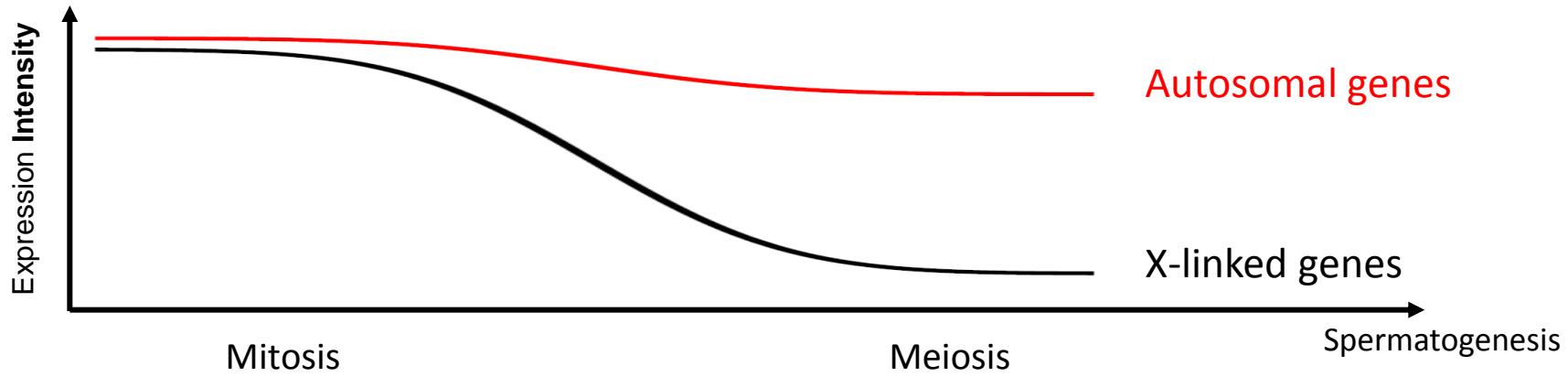


4. Statistical Approaches

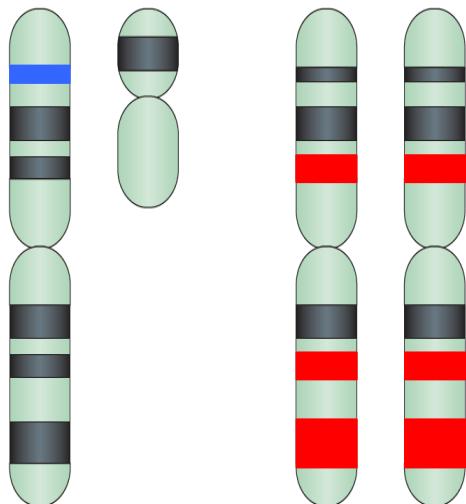
Meiotic Sex Chromosome Inactivation (MSCI)



2. Biological Problem



2. Biological Problem

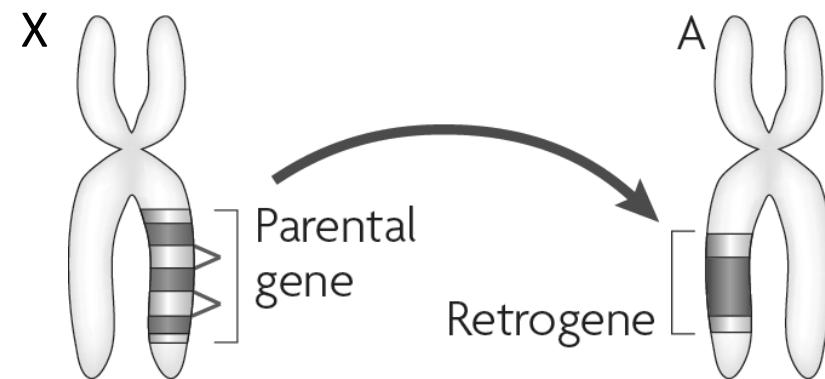


X Y X X



XY systems

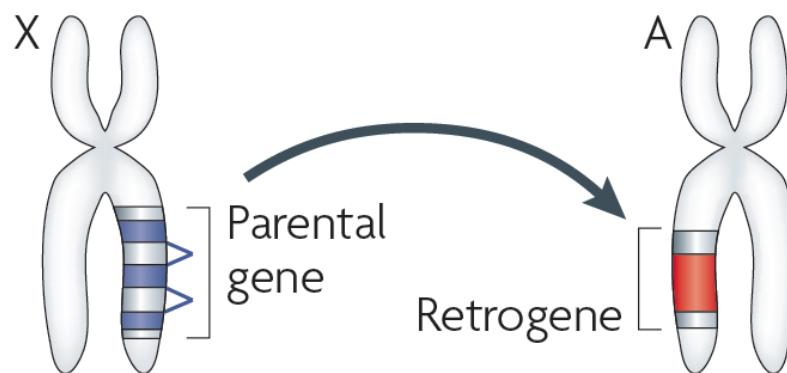
- Does MSCI exist in *Drosophila*? YES
- Is MSCI involved in the distribution of sex-biased genes in the genome?



Testis bias

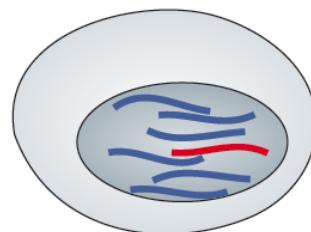
2. Biological Problem

Out-of-the-X retroposition

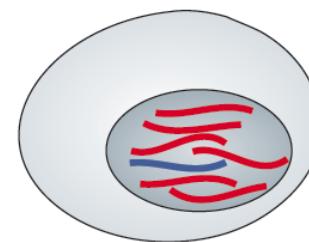


Parental gene and retrogene transcription levels
during distinct stages of
spermatogenesis

- Parental gene mRNA
- Retrogene mRNA



Spermatogonia
(Mitotic)



Spermatocytes
(Meiotic)

Intensity of sex-chromosome
inactivation during and after
meiosis

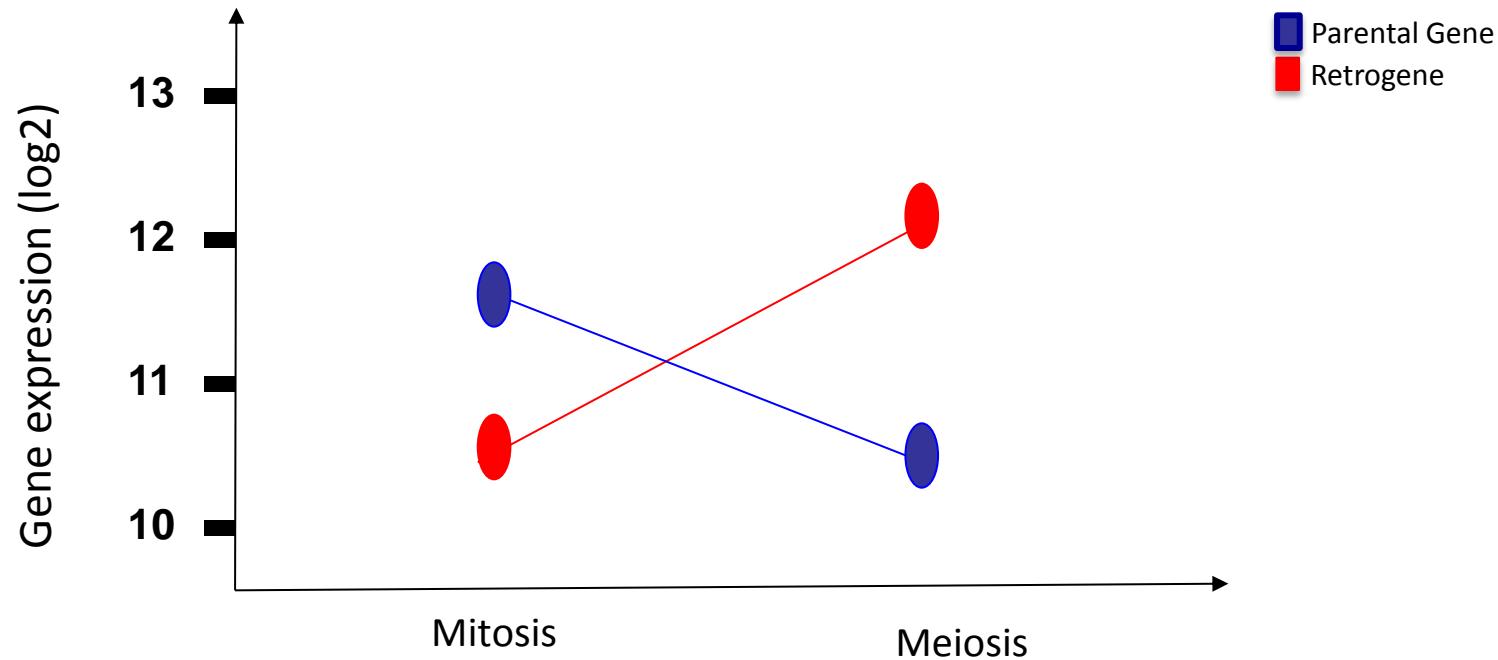


2. Biological Problem

Complementary expression

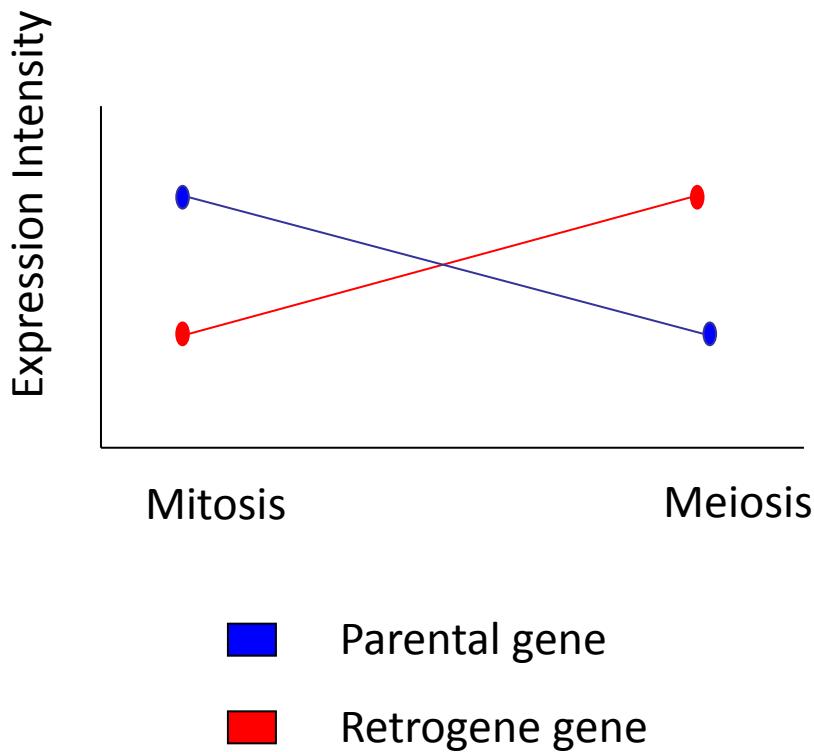
Tom40: Translocase of outer membrane 40 / Chr X

Tomboy40: protein transmembrane transporter activity / Chr 2R

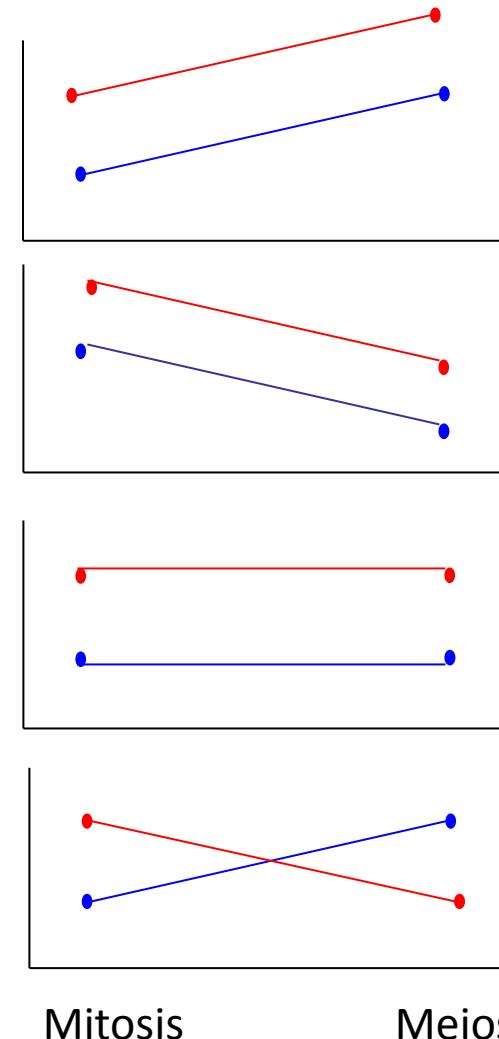


2. Biological Problem

Complementary expression

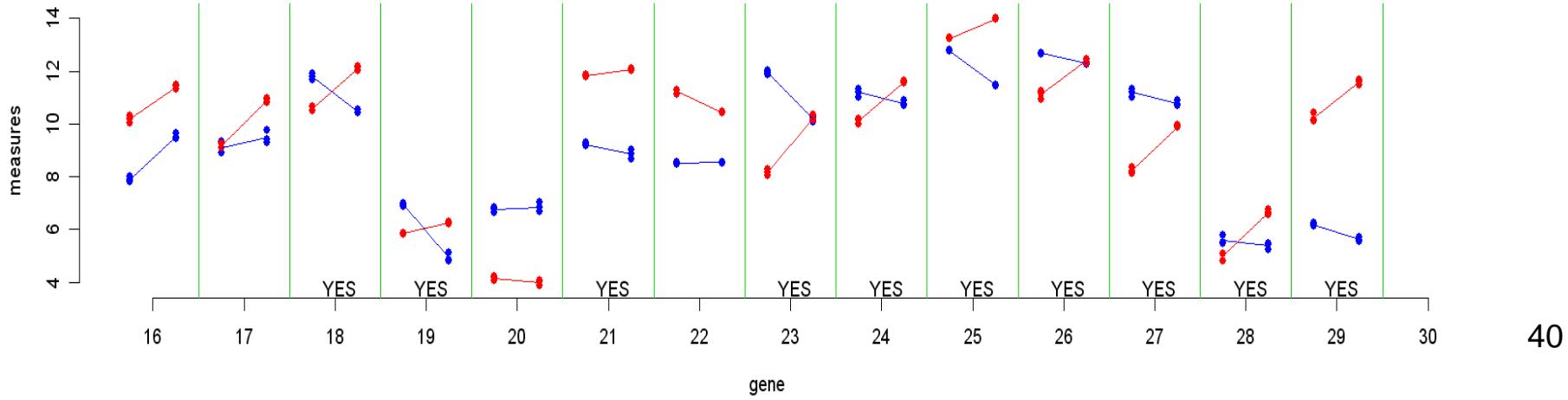
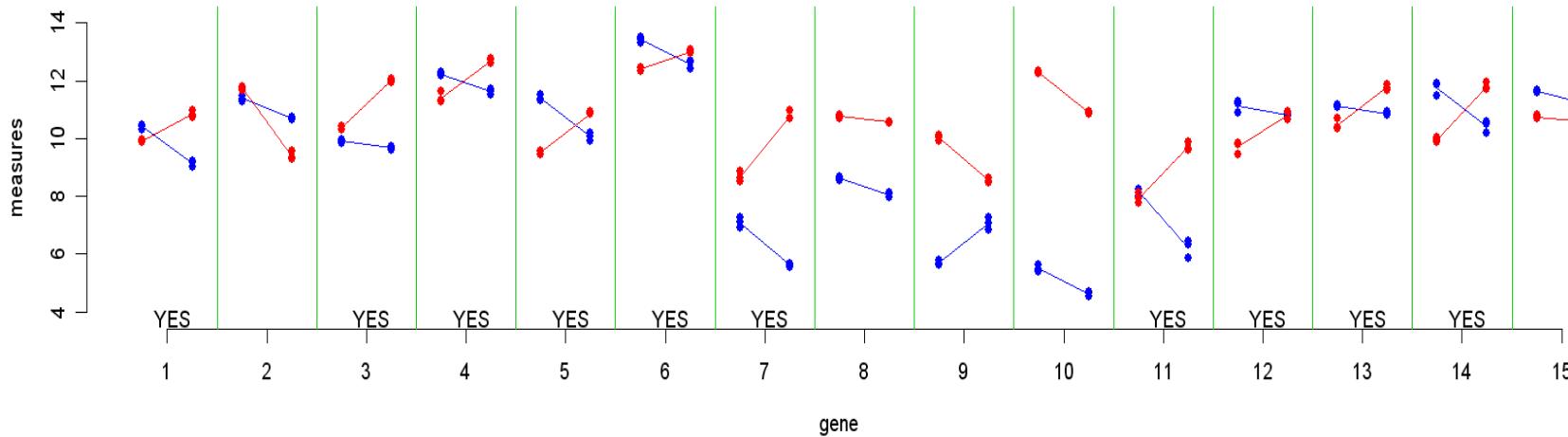


No Complementary expression



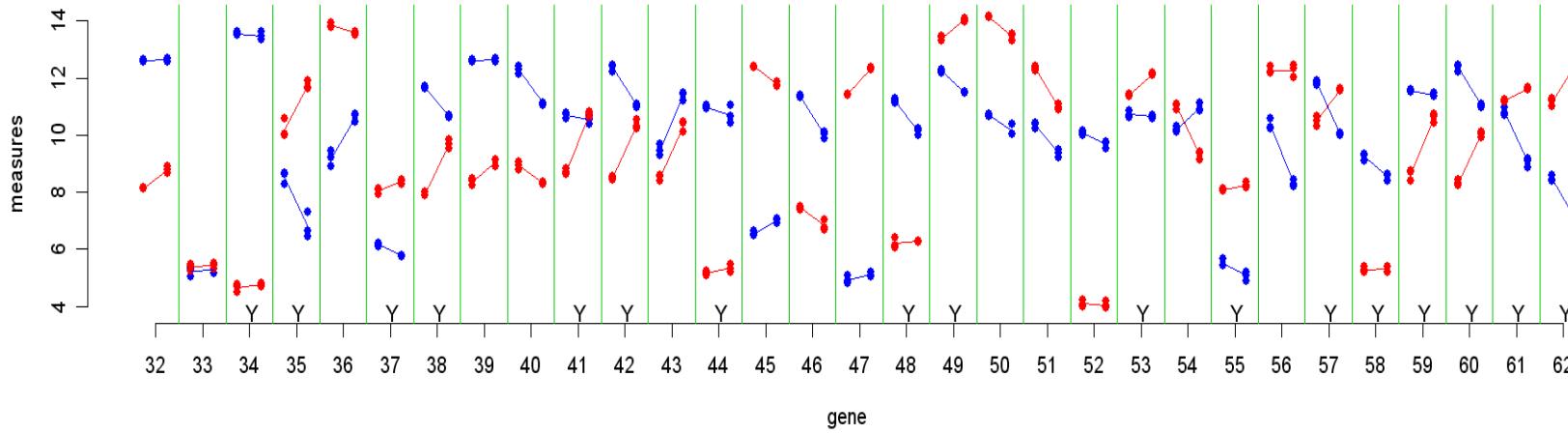
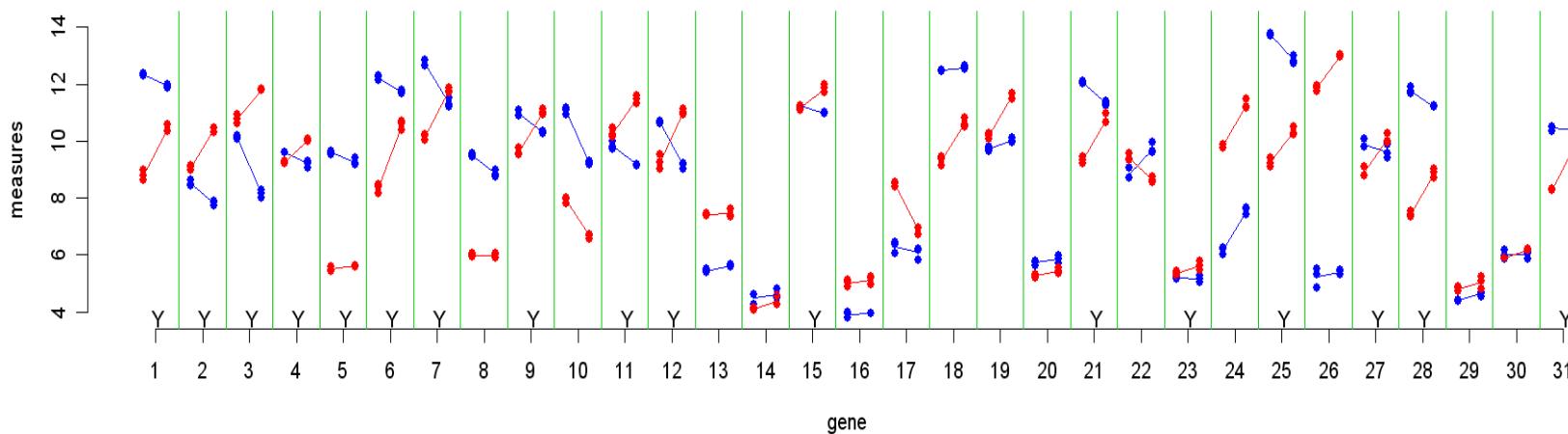
4. Statistical Approaches

Out of X



4. Statistical Approaches

Out of A



4. Statistical Approaches

Approach 1: Counting YES and NOS

Counts	NO	YES	Proportions	NO	YES
Out of X	9	20	Out of X	0.31	0.69
Out of A	28	34	Out of A	0.45	0.55

Fisher's Exact Test for Count Data

Null hypothesis: true odds ratio = 1

Alternative hypothesis: true odds ratio is not equal to 1

p-value = 0.2546

Estimated odds ratio=0.55 (roughly $(34/28)/(20/9)=0.546$)

95% confidence interval: (0.1888,1.5086)

95% confidence interval for YES (normal approximation)

Out of X: (0.518,0.862)

Out of A: (0.424,0.676)

4. Statistical Approaches

Approach 2: Controlling for FDR

Considering only significant change of expression 2 fold difference $p < 0.05$, $q < 0.05$

Counts	NO	YES	Proportions	NO	YES
Out of X	9	18	Out of X	0.33	0.66
Out of A	37	25	Out of A	0.60	0.40

Fisher's Exact Test for Count Data

Null hypothesis: true odds ratio = 1

Alternative hypothesis: true odds ratio is not equal to 1

p-value = 0.03687

Estimated odds ratio=0.342 (roughly $(25/37)/(18/9)=0.339$)

95% confidence interval: (0.11,0.95)

95% confidence interval for YES (normal approximation)

Out of X: (0.478,0.842)

Out of A: (0.276,0.524)

4. Statistical Approaches

Approach 3: Bayesian hierarchical model

$$\begin{pmatrix} mit_{ijkl} \\ mei_{ijkl} \end{pmatrix} \sim N \left[\begin{pmatrix} \theta_{ijl}^{mit} \\ \theta_{ijl}^{mei} \end{pmatrix}, \sigma_i^2 I_2 \right]$$

$$\begin{pmatrix} \theta_{ijl}^{mit} \\ \theta_{ijl}^{mei} \end{pmatrix} \sim N \left[\begin{pmatrix} \theta_{ij}^{mit} \\ \theta_{ij}^{mei} \end{pmatrix}, \begin{pmatrix} \tau_{mit}^2 & 0 \\ 0 & \tau_{mei}^2 \end{pmatrix} \right]$$

Pairs mit_{ijkl} and mei_{ijkl} , for each gene l , each classification group i (out of X, out of A) and gene type j (parental, offspring), have individual means θ_{ijl}^{mit} and θ_{ijl}^{mei} , respectively, and common classification group variances σ_i^2 . Then, the objective is to compute,

$$\Pr \left[\text{YES}_{\text{Out of X}, l} \right] = \Pr \left[\left\{ \theta_{\text{Out of X, par}, l}^{mit} > \theta_{\text{Out of X, par}, l}^{mei} \right\} \cap \left\{ \theta_{\text{Out of X, off}, l}^{mit} < \theta_{\text{Out of X, off}, l}^{mei} \right\} \right]$$

$$\Pr \left[\text{YES}_{\text{Out of A}, l} \right] = \Pr \left[\left\{ \theta_{\text{Out of A, par}, l}^{mit} > \theta_{\text{Out of A, par}, l}^{mei} \right\} \cap \left\{ \theta_{\text{Out of A, off}, l}^{mit} < \theta_{\text{Out of A, off}, l}^{mei} \right\} \right]$$

4. Statistical Approaches

Approach 3: Bayesian hierarchical model

Hyperparameters

$$\Theta = (\theta_{11}^{mit}, \theta_{12}^{mit}, \theta_{21}^{mit}, \theta_{22}^{mit}, \theta_{11}^{mei}, \theta_{12}^{mei}, \theta_{21}^{mei}, \theta_{22}^{mei})$$

$$\Lambda = (\sigma_1^2, \sigma_2^2, \tau_{mit}^2, \tau_{mei}^2)$$

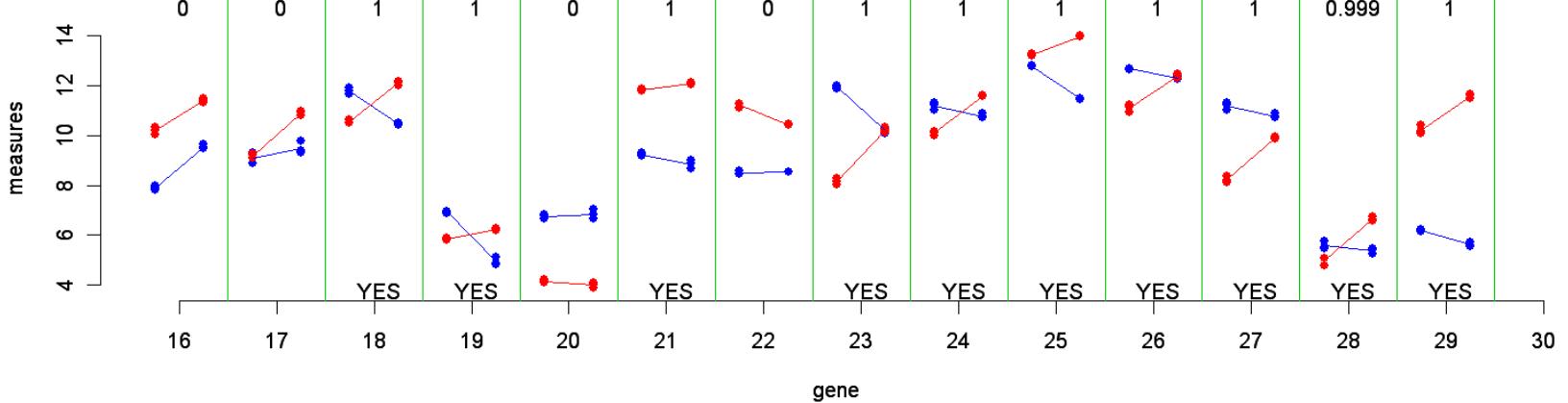
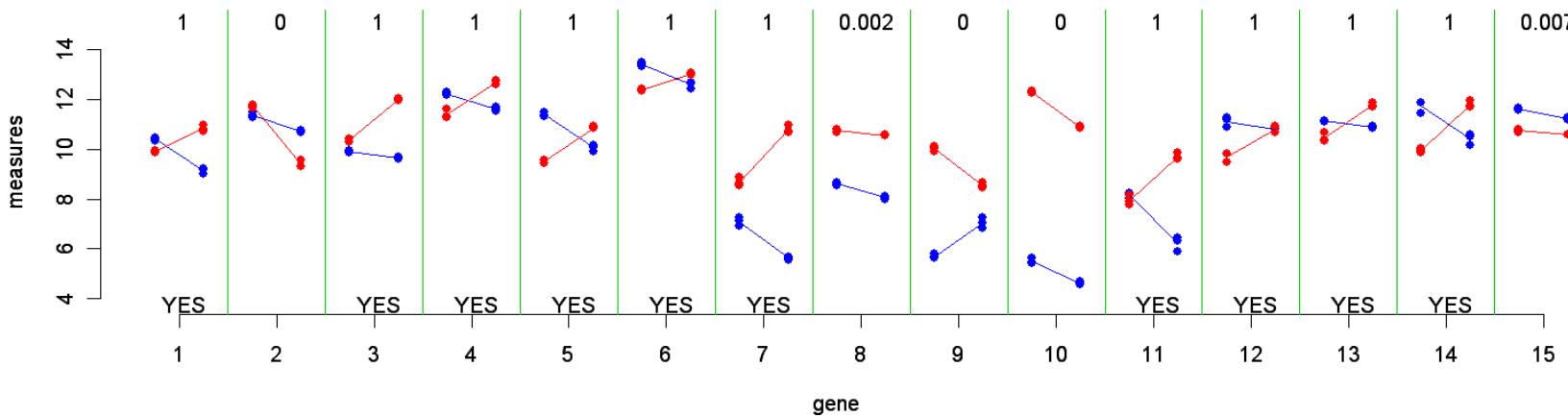
Prior distribution

$$p(\Theta, \Lambda) \propto \sigma_1^{-2} \sigma_2^{-2} \tau_{mit}^{-2} \tau_{mei}^{-2}$$

This represents vague/noninformative prior views.

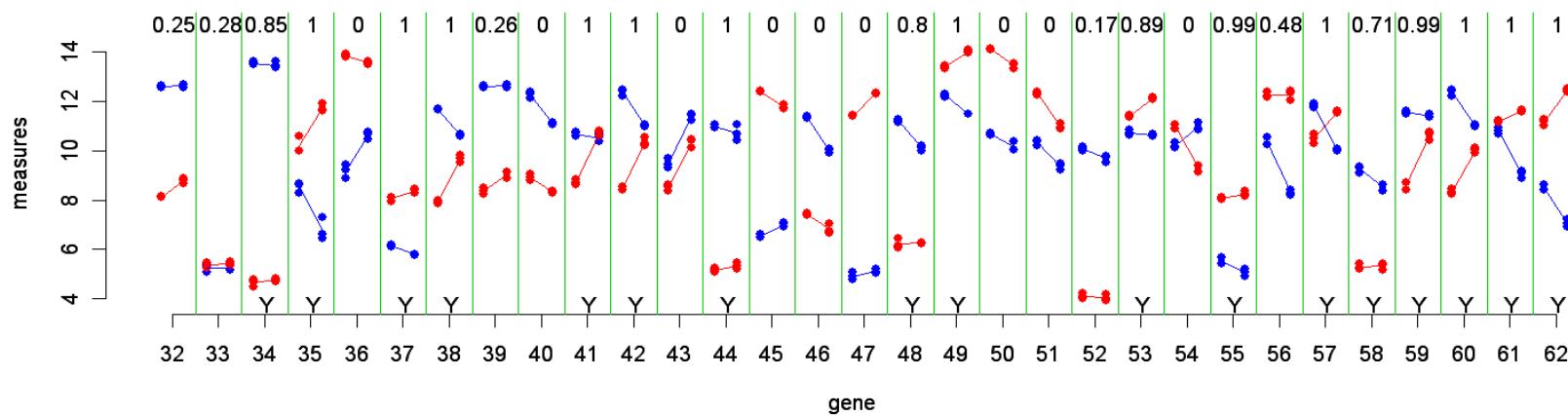
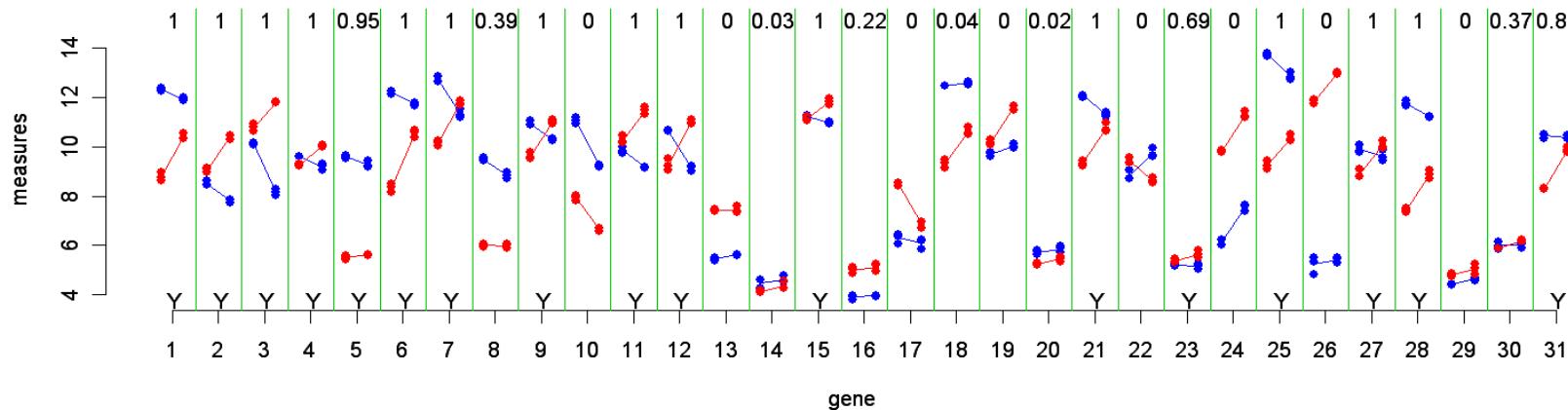
4. Statistical Approaches

Approach 3: Bayesian hierarchical model

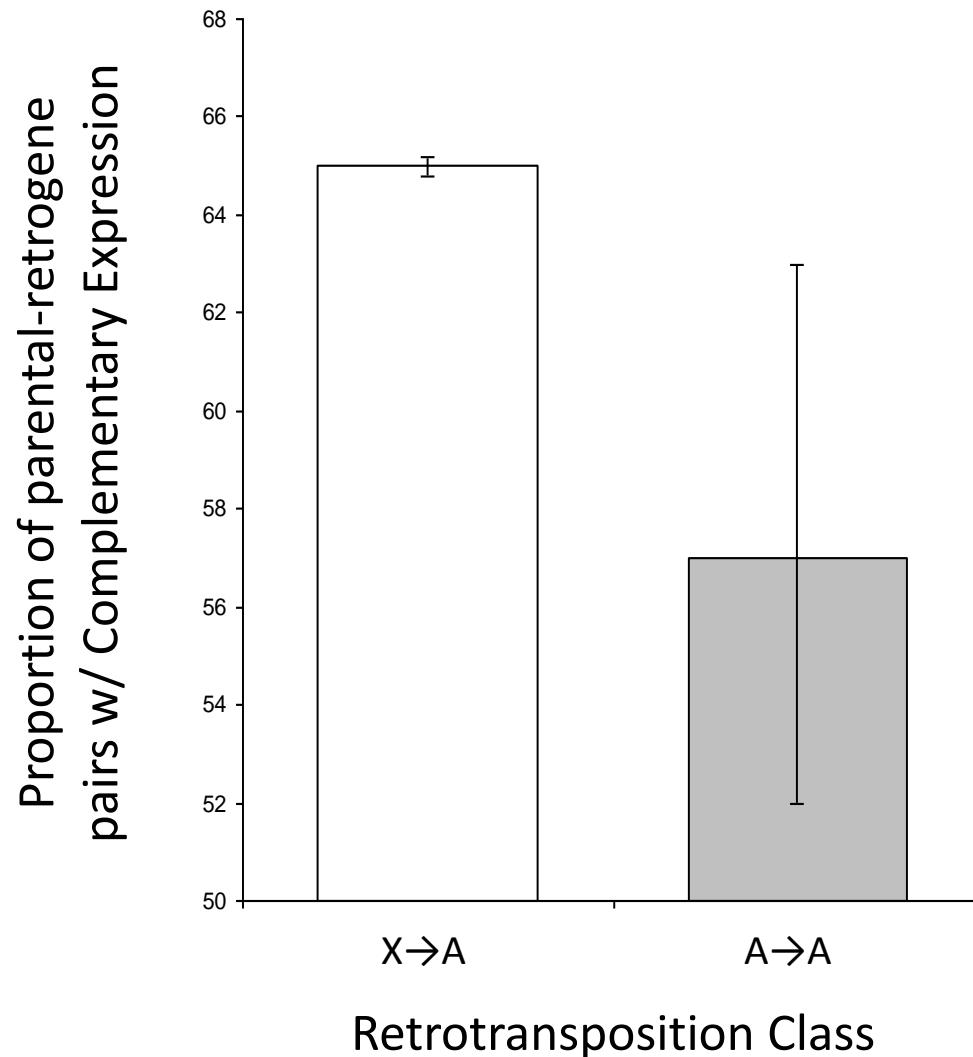


4. Statistical Approaches

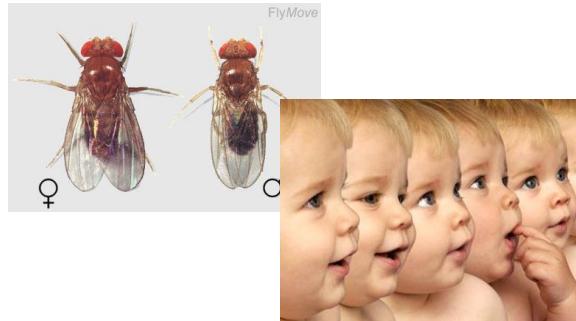
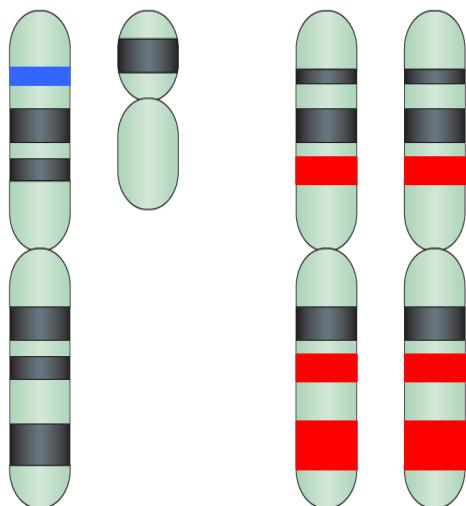
Approach 3: Bayesian hierarchical model



4. Statistical Approaches

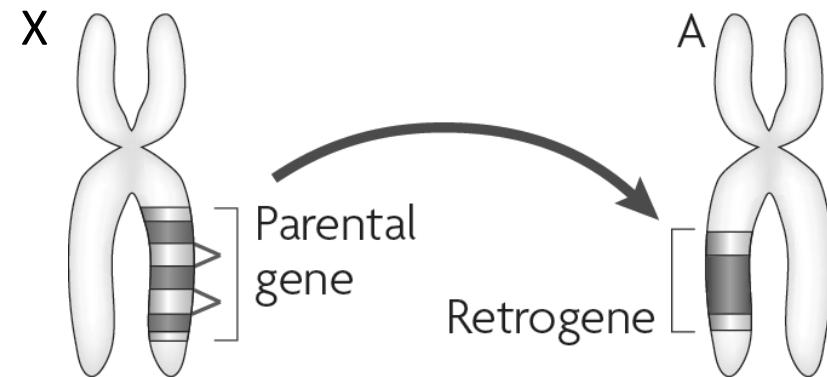


2. Biological Problem



XY systems

- Does MSCI exist in *Drosophila*? YES
- Is MSCI involved in the distribution of sex-biased genes in the genome? YES



Testis bias

2. Biological Problem

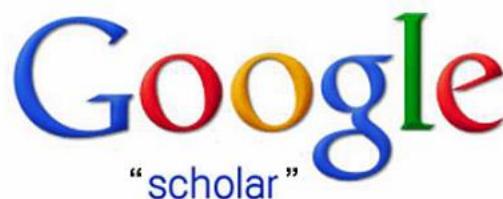
OPEN  ACCESS Freely available online

PLOS GENETICS

Stage-Specific Expression Profiling of *Drosophila* Spermatogenesis Suggests that Meiotic Sex Chromosome Inactivation Drives Genomic Relocation of Testis-Expressed Genes

Maria D. Vibranovski¹, Hedibert F. Lopes², Timothy L. Karr³, Manyuan Long^{1*}

¹ Department of Ecology and Evolution, The University of Chicago, Chicago, Illinois, United States of America, ² The University of Chicago Booth School of Business, Chicago, Illinois, United States of America, ³ The Biodesign Institute, Arizona State University, Tempe, Arizona, United States of America



Título 1–20

Stage-specific expression profiling of *Drosophila* spermatogenesis suggests that meiotic sex chromosome inactivation drives genomic relocation of testis-expressed genes

MD Vibranovski, HF Lopes, TL Karr, M Long
PLoS genetics 5 (11), e1000731

Citado por 109

Ano 2009

2. Biological Problem

Male infertility



In Humans:

- 40% of infertility
- High cost treatment
- Associated with gene expression/function in spermatogenesis
- 30% of infertility are sperm deficiency

Drosophila Model:

- Spermatozoa is one of the few cell types that has homologous function for all sexual organism including humans
- Post-meiotic transcription

2. Biological Problem

SpermPress


Drosophila spermatogenesis expression database

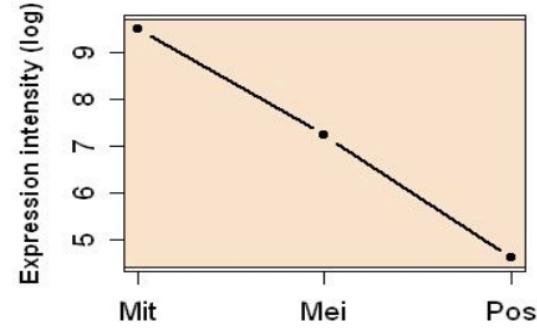
Search for CG or Affymetrix gene symbol/probe id Browse gene list

CG id: CG10422
Affymetrix gene: bam
Affymetrix probe id: 1634336_at

Gene expression levels

phase	mitosis	meiosis	post-meiosis
value 1	9.32	6.94	4.61
value 2	9.64	7.37	4.72
value 3	9.51	7.38	4.56
average	9.49	7.23	4.63

Values 1-3 correspond to the expression intensity (log transformed) obtained from Microarray hybridization of three different biological replicates of each spermatogenic phase. Averages values correspond to the arithmetic average within replicates for each spermatogenic phase.



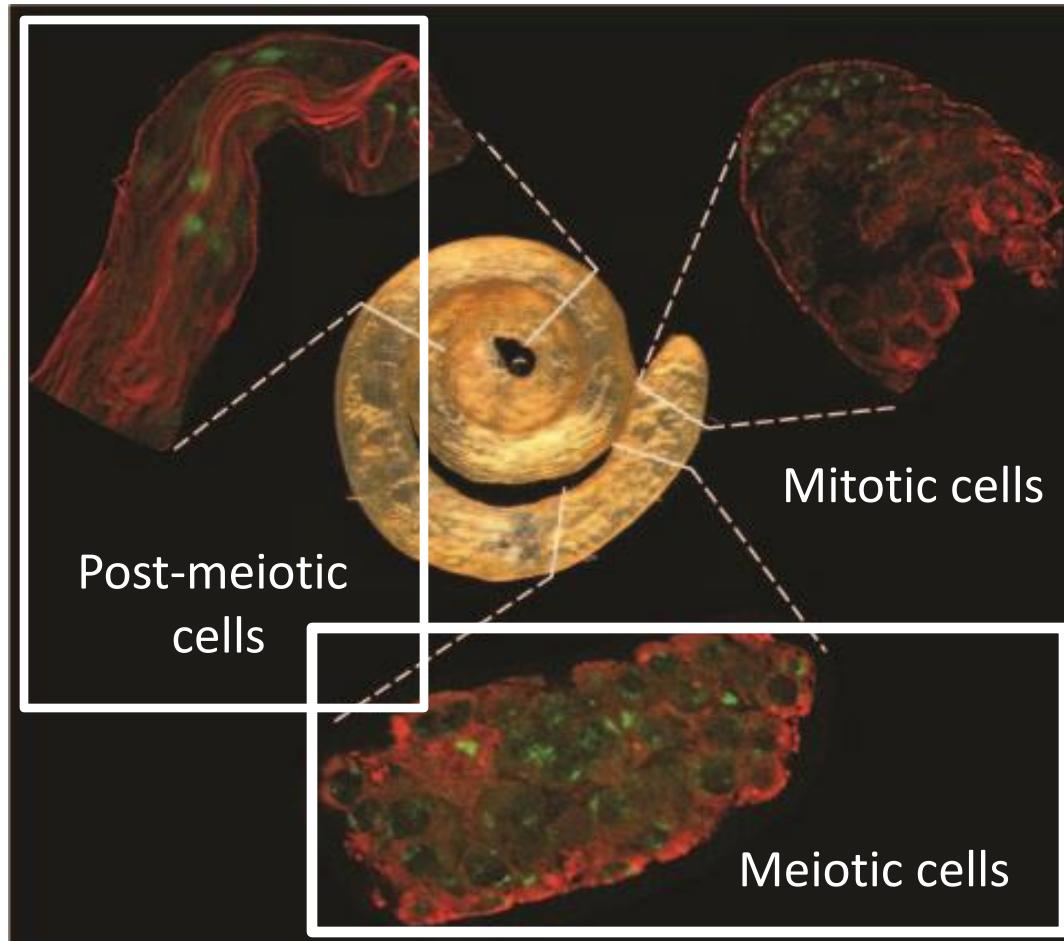
phase	mitosis	meiosis	post-meiosis
mitosis	-	Over	Over
meiosis	-	-	Over
post-meiosis	-	-	-

Comparison of expression levels

phase	mitosis	meiosis	post-meiosis
mitosis	-	Over	Over
meiosis	-	-	Over
post-meiosis	-	-	-

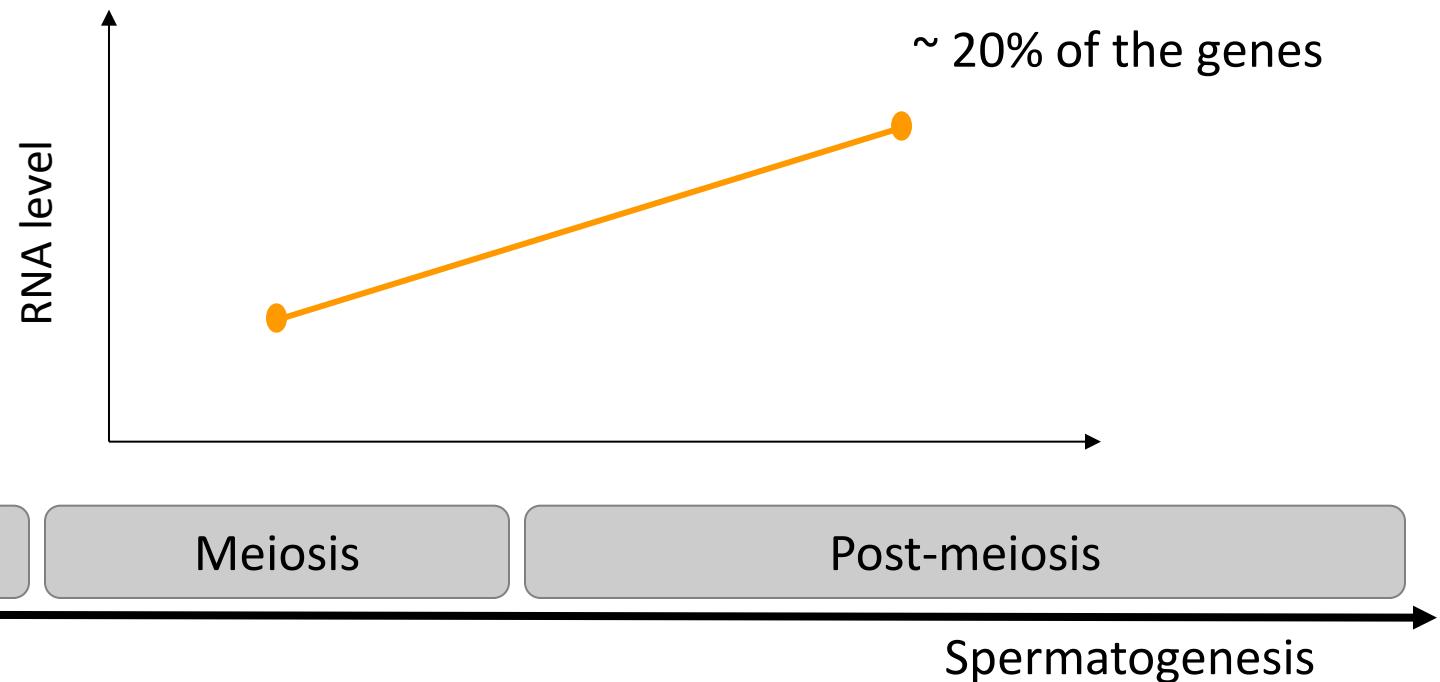
The classifications in this table (Equal, Over or Under) were obtained by our Bayesian Statistical Model (Methods). Each classification is given to a pair-wise comparison between two spermatogenic phases. For example, "Over" mitotic vs. meiotic classification means that, for this gene, the mitotic expression is significantly higher than the meiotic expression.

5. Ongoing Biological Problems



5. Ongoing Biological Problems

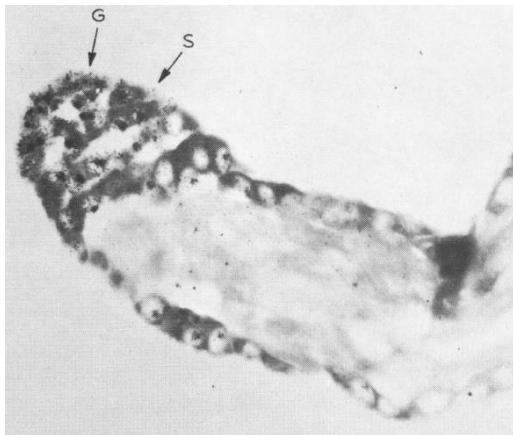
Post-meiosis Over-expression



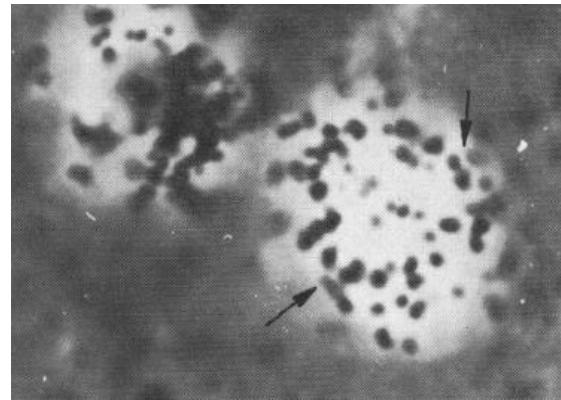
5. Ongoing Biological Problems

RNA synthesis does **NOT** occur in post-meiotic stages

Testis Apical region

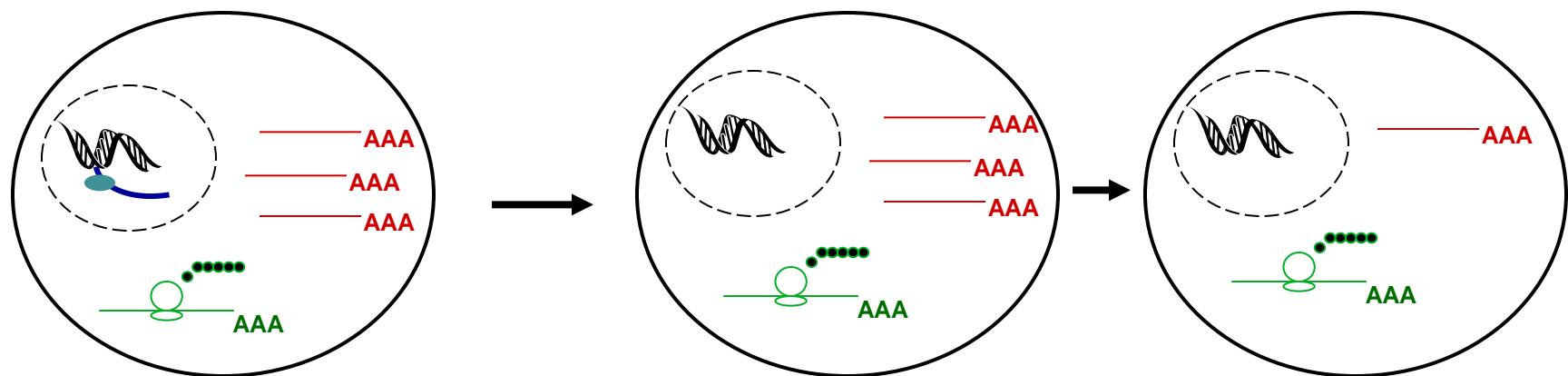


Spermatocytes



RNA synthesis: incorporation of [³H]uridine in the *Drosophila melanogaster* testes has been studied by autoradiography.

5. Ongoing Biological Problems



Transcription

Translation

RNA

Mitosis

Meiosis

Post-meiosis

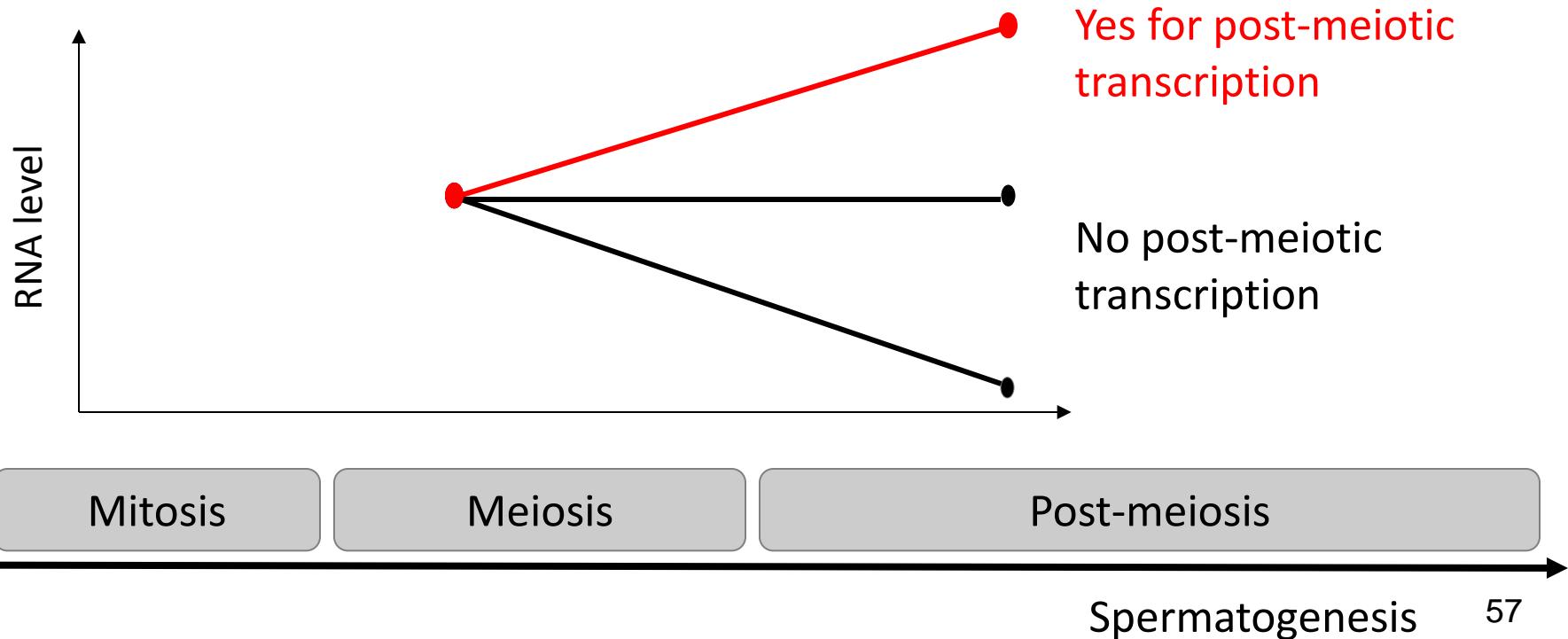
Spermatogenesis

5. Ongoing Biological Problems

Transcription

Translation

RNA



Mitosis

Meiosis

Post-meiosis

Spermatogenesis

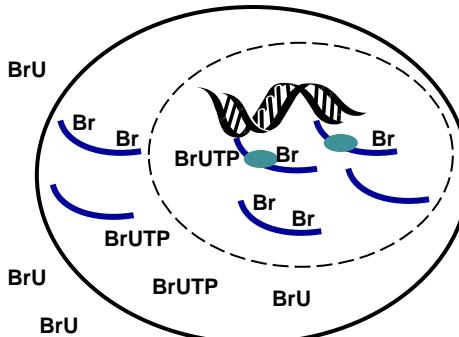
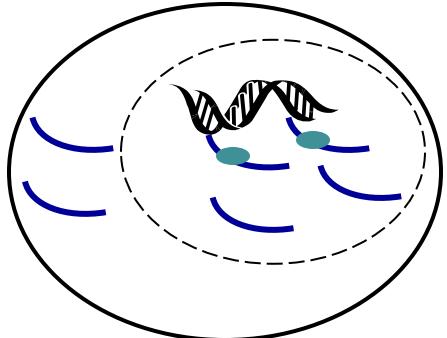
57

5. Ongoing Biological Problems

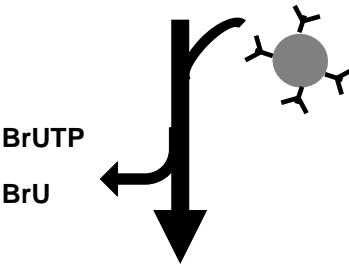
Bromo-uridine (BrU) incorporation: Direct evidence



Domitille Chalopin

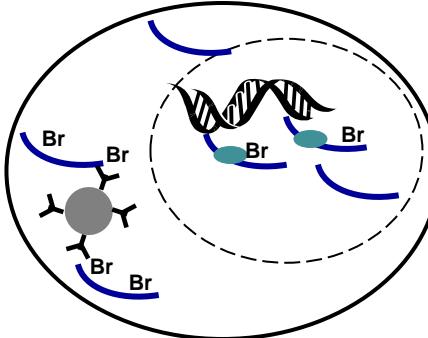
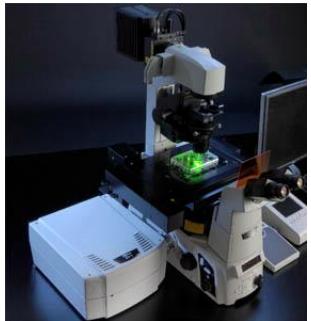


- (1) Incorporation of BrU
- (2) Labeling nascent RNAs with Br-UTP



- (3) Binding anti-BrdU antibody

←
(4) Imaging

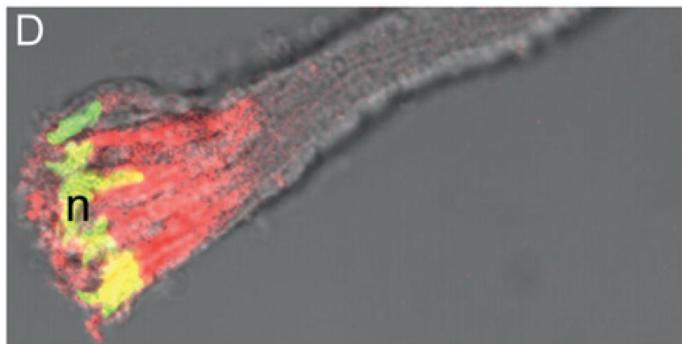


5. Ongoing Biological Problems

Bromo-uridine (BrU) incorporation:
Post-meiotic transcription - Direct evidence



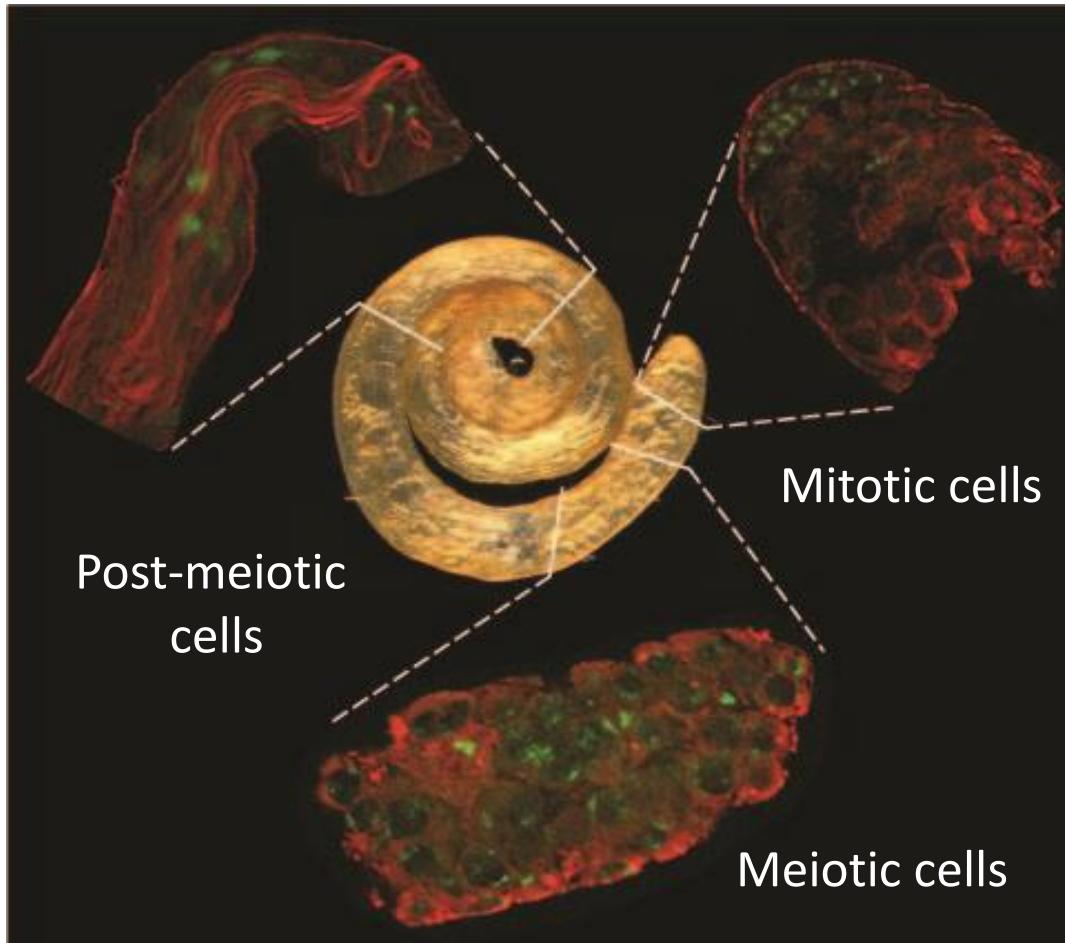
Domitille Chalopin



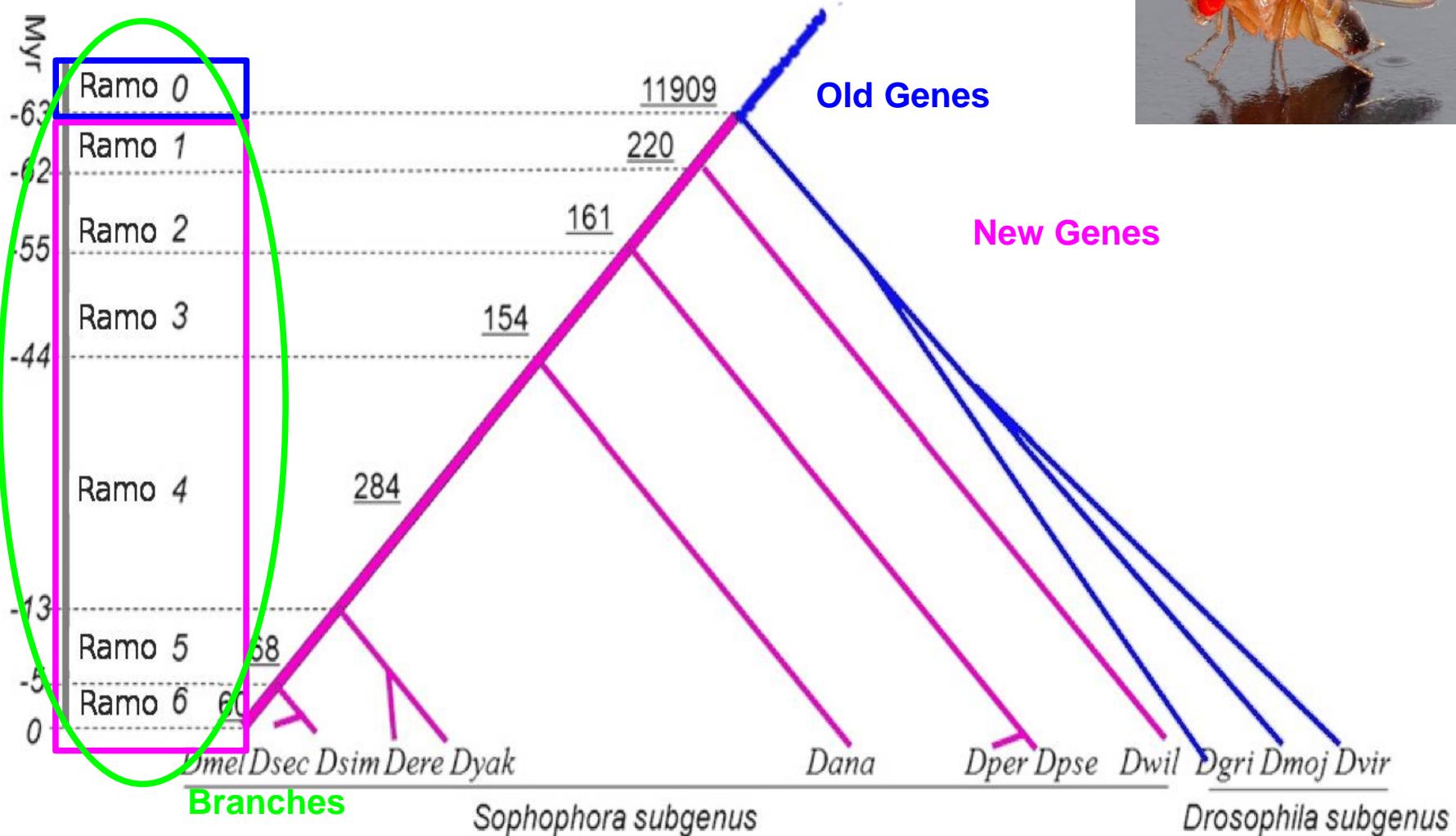
BrU
DNA
n: nuclei

Spermatid cyst

5. Ongoing Biological Problems



5. Ongoing Biological Problems

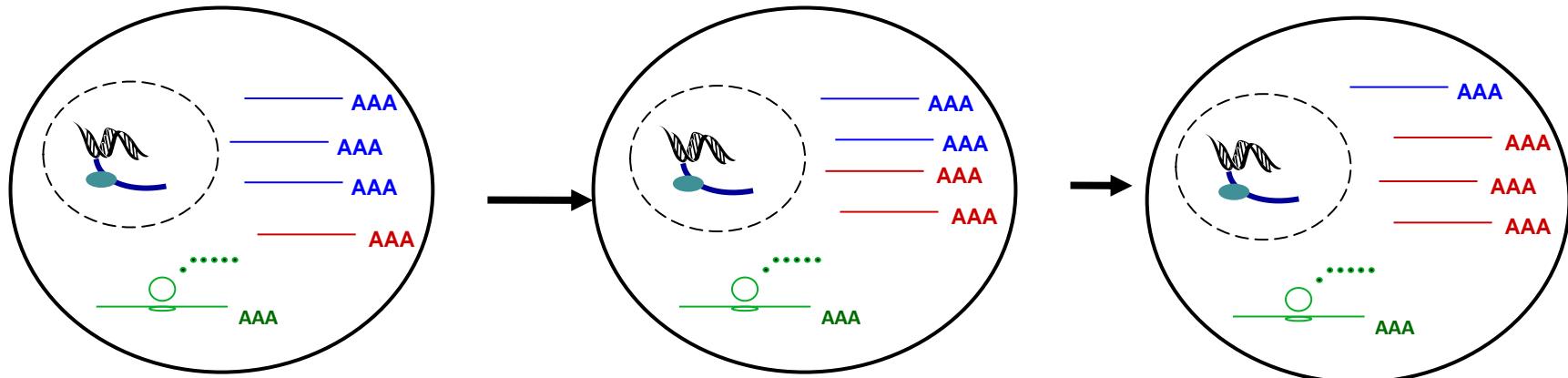


Zhang, Vibranovski, Krinsky and Long, 2010

5. Ongoing Biological Problems



Júlia Raices



New genes RNA

Old genes RNA

Mitosis

Meiosis

Post-meiosis

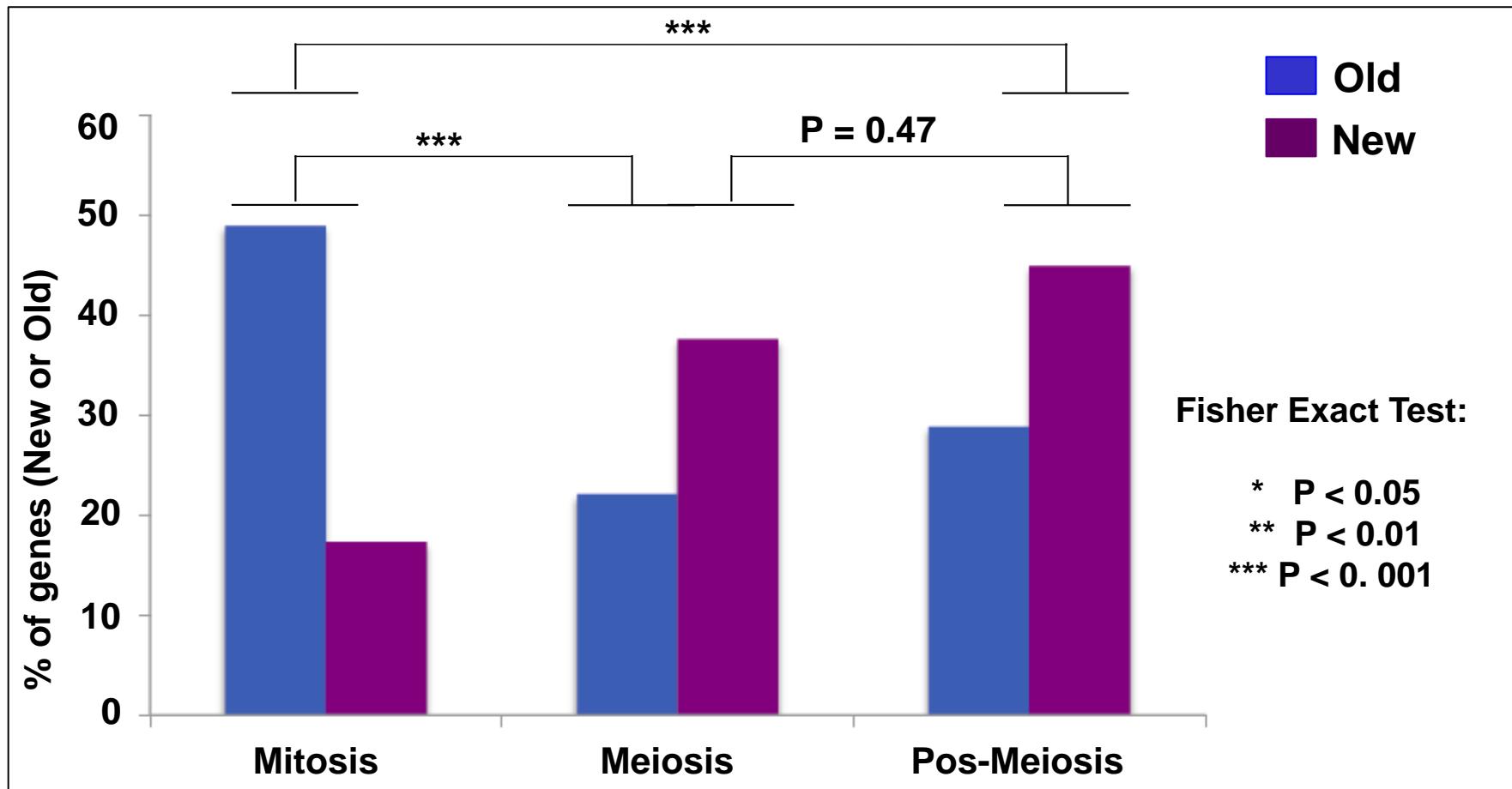
Spermatogenesis

5. Ongoing Biological Problems



Júlia Raices

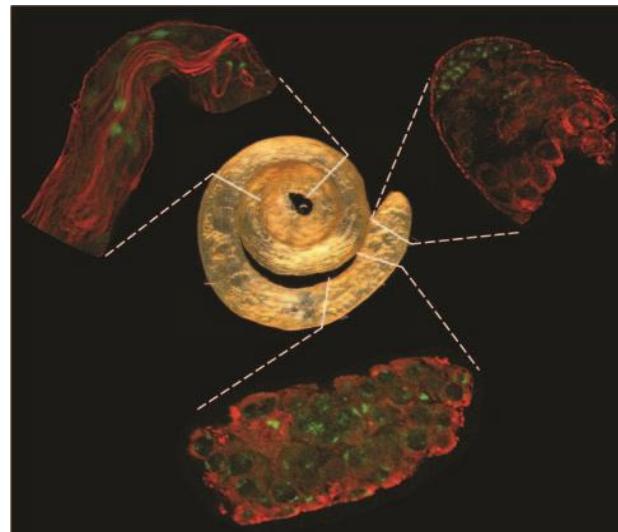
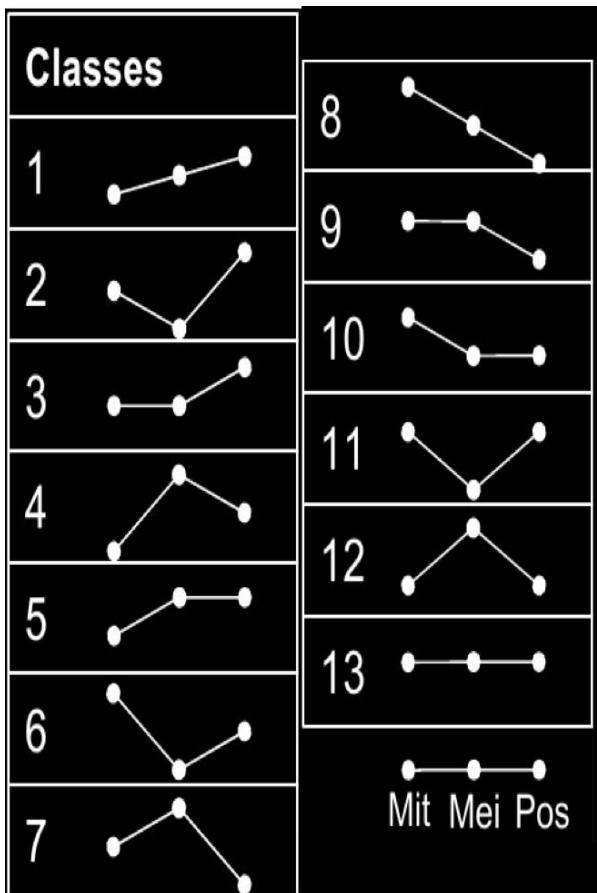
Proportion of Genes by Spermatogenesis Phase and by age



5. Ongoing Biological Problems



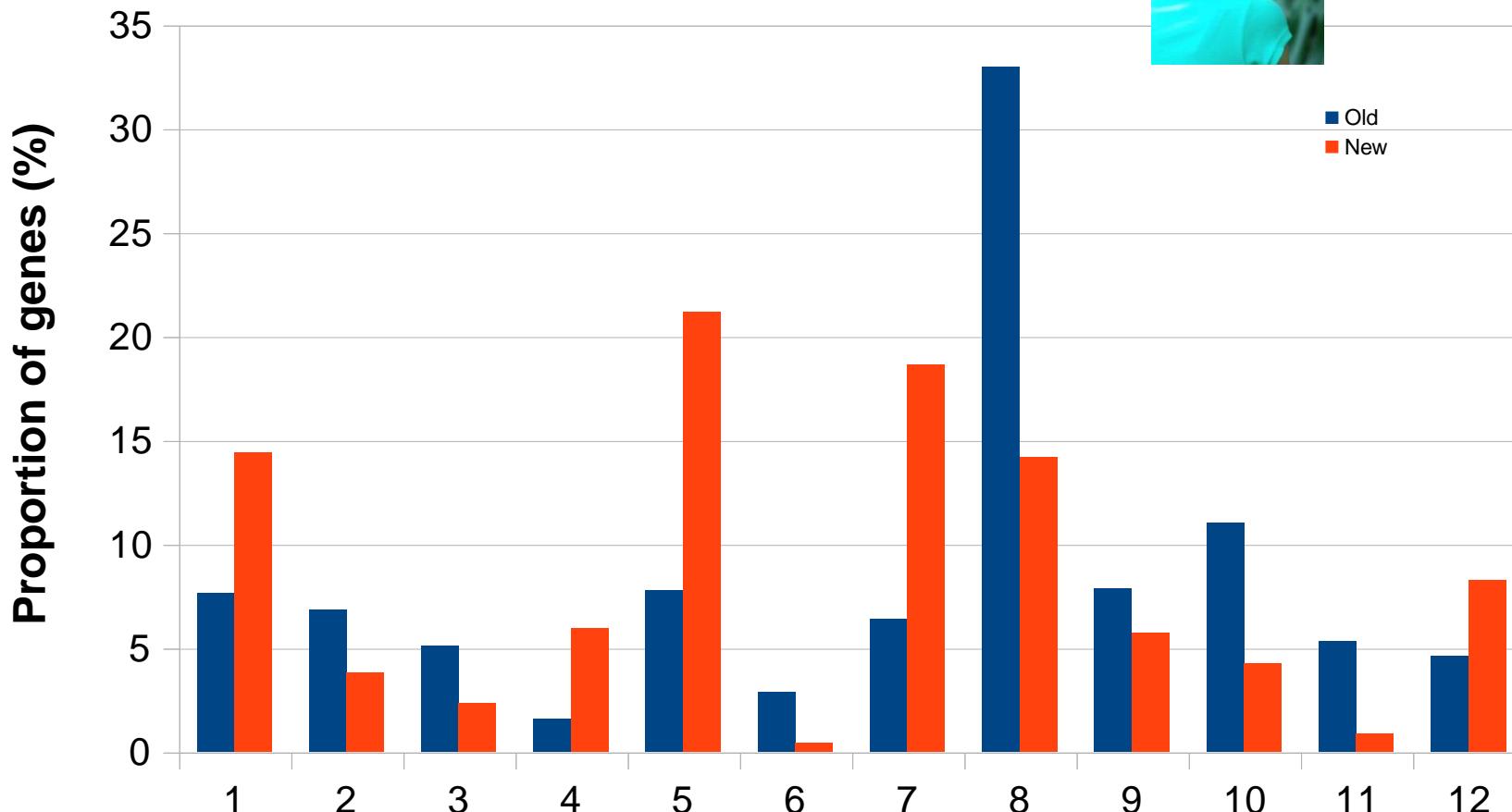
Júlia Raices



5. Ongoing Biological Problems

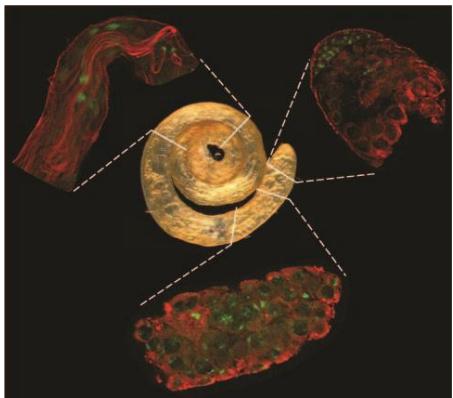


Júlia Raices



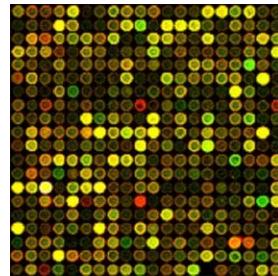
Perspectives

Development

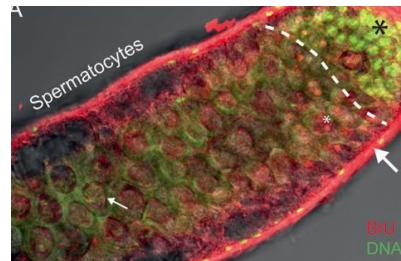


Genetics

Molecular Biology



Cell Biology



Bioinformatics



Statistics

Acknowledgments

THE VIBRANOVSKI LAB



Gustavo França
IQ /USP



Júlia Raices
IB /USP



Mariana Kanbe
ICB /USP



Joana Carvalho
IB /USP

THE LONG LAB



Manyuan Long
Yuan Huang,
Claus Kemkemer,
Ben Krinsky
Patrick Landback
Nick Vankuren
Jun Wang
Qian Yang
Chengjung Zhang

Post-meiotic transcription

Domitille Chalopin, ENS, Lyon

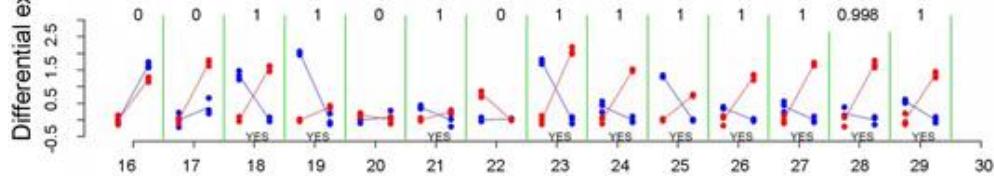
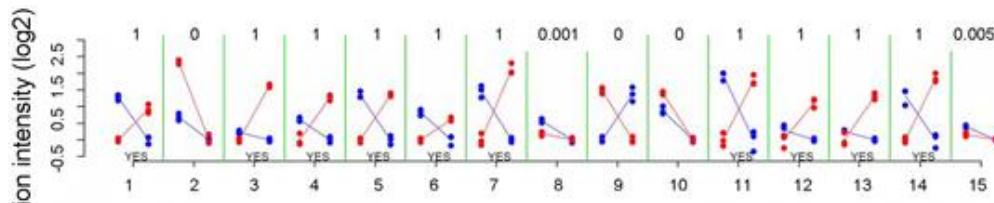
Hedibert Lopes

Timothy Karr



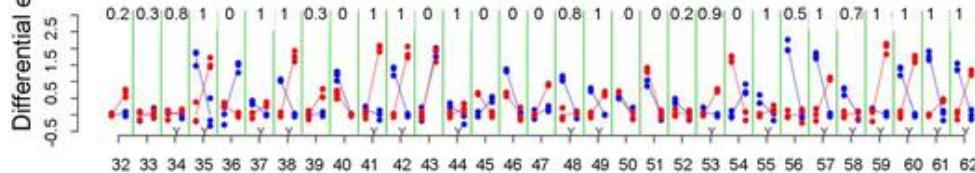
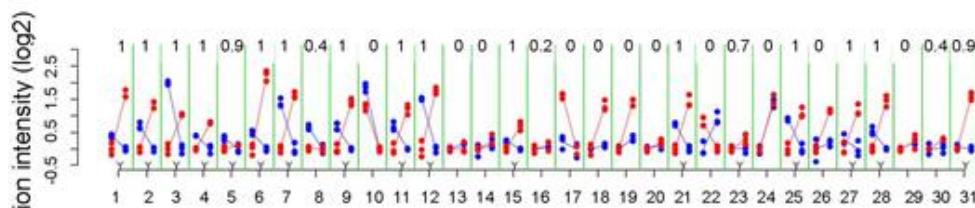
Funding:





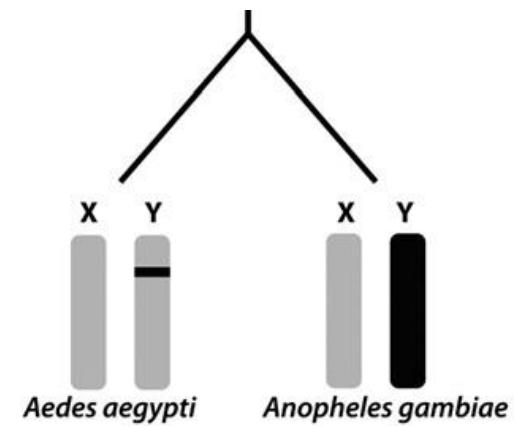
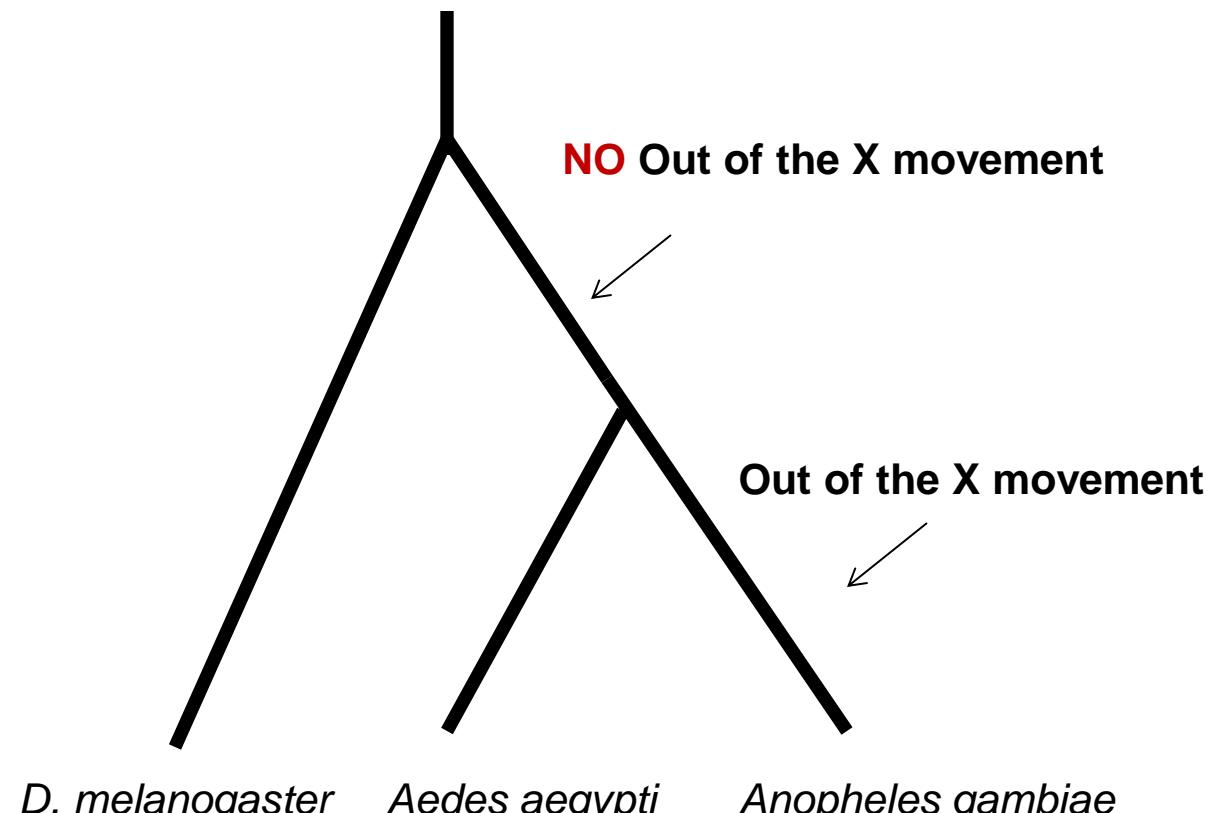
X → A

A



A(A)

B



Toups and Hahn, 2010, Genetics